

VIASURE

Real Time PCR Detection Kits

by CerTest
BIOTEC

SARS-CoV-2 (N1 + N2)

Handbook for the following references/

Priručnik za sljedeće reference:

VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System

BD REF 444215

to be used with the BD MAX™ System

koristi se sa BD MAX™ sustavom



ENGLISH

1. Intended use

VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System is an automated real-time RT-PCR test designed for the qualitative detection of RNA from the SARS-CoV-2 in respiratory samples from individuals suspected of COVID-19 by their healthcare provider. This test is intended to be used as an aid in the identification of the presence of the SARS-CoV-2 viral RNA. The assay uses the BD MAX™ System for automated extraction of RNA and subsequent real-time RT-PCR employing the reagents provided combined with universal reagents and disposables for the BD MAX™ System. RNA is extracted from respiratory specimens, amplified using RT-PCR and detected using fluorescent reporter dye probes specific for SARS-CoV-2.

2. Summary and Explanation

Coronavirus are enveloped non-segmented positive-sense RNA viruses and belong to Coronaviridae family [1,2]. There are six coronavirus species known to cause human diseases [2]. Four viruses (229E, OC43, NL63 and HKU1) cause common cold symptoms and the other two (severe acute respiratory syndrome coronavirus (SARS-CoV) and Middle East respiratory syndrome coronavirus (MERS-CoV)) are zoonotic and producing more severe complications [2]. SARS-CoV and MERS-CoV have caused more than 10,000 cumulative cases in the past two decades, with mortality rates of 34% MERS-CoV and 10% SARS-CoV [1,3].

In December 2019, some people that worked at or lived around the Huanan seafood market in Wuhan, Hubei Province, China, have presented pneumonia of unknown cause [2,4]. Deep sequencing analysis of the respiratory samples indicated a novel coronavirus, which was named firstly 2019 novel coronavirus (2019-nCoV) and lately SARS-CoV-2 [5].

Human-to-human transmission of the SARS-CoV-2 has been confirmed, even in the incubation period without symptoms, and the virus causes severe respiratory illness like those SARS-CoV produced [1,6,7,8]. Although the pneumonia is the principal illness associated, a few patients have developed severe pneumonia, pulmonary edema, acute respiratory distress syndrome, or multiple organ failure and death [1,4]. Centers of Disease Control and Prevention (CDC) believes that symptoms of SARS-CoV-2 may appear in as few as 2 days or as long as 14 days after exposure, being the most common fever or chills, cough, fatigue, anorexia, myalgia and dyspnea [1,4,6,9]. Less common symptoms are sore throat, nasal congestion, headache, diarrhea, nausea and vomiting [1,4]. Loss of smell (anosmia) or loss of taste (ageusia) preceding the onset of respiratory symptoms has also been reported [9]. Older adults and people who have severe underlying medical conditions like heart or lung disease or diabetes seem to be at higher risk for developing more serious complications from COVID-19 illness [10].

Diagnosis of SARS-CoV-2 is performed detecting conventional causes of pneumonia early and detected by next-generation sequencing or real-time RT-PCR methods [1,11]. Several assays that detect the SARS-CoV-2 have been currently available, such as China CDC (gene targets, ORF1ab and N), Charité – Germany (gene targets, RdRP and E) or US CDC (two targets in N gene) [12].

CDC recommends upper respiratory tract specimens (nasopharyngeal (NP) and oropharyngeal (OP) swabs, nasal mid-turbinate swab, nasal swab, nasopharyngeal wash/aspirate or nasal wash/aspirate (NW) specimens collected



mainly by a healthcare provider) and/or lower respiratory specimens (sputum, endotracheal aspirate, or bronchoalveolar lavage in patients with more severe respiratory disease) for the identification of SARS-CoV-2 [11]. In addition, other clinical specimens as blood, urine and stool may be collected to monitor the presence of the virus [11,12].

3. Principle of the procedure

VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System is designed for the identification of SARS-CoV-2 in respiratory samples. The detection is done in one step real-time RT-PCR format where the reverse transcription and the subsequent amplification of specific target sequence occur in the same reaction tube. The isolated RNA target is transcribed generating complementary DNA by reverse transcriptase which is followed by the amplification of two conserved regions of N gene (N1 and N2) using specific primers and fluorescent-labeled probes.

VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System is based on 5' exonuclease activity of DNA polymerase. During DNA amplification, this enzyme cleaves the probe bound to the complementary DNA sequence, separating the quencher dye from the reporter. This reaction generates an increase in the fluorescent signal which is proportional to the quantity of the target template. This fluorescence is measured on the BD MAX™ System.

VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System contains in each tube all the components necessary for real-time PCR assay (specific primers/probes, dNTPS, buffer, polymerase, reverse-transcriptase) in a stabilized format, as well as an endogenous internal control to monitor the extraction process and/or inhibition of the polymerase activity. The assay uses a human housekeeping gene as an endogenous Internal Control (IC) (human RNase P gene). Human housekeeping genes are involved in basic cell maintenance and, therefore, are expected to be present in all nucleated human cells and maintain relatively constant expression levels. N2 target is amplified and detected in channel 475/520, N1 target in channel 630/665 and the endogenous internal control (IC) in channel 530/565.

4. Reagents provided

VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System includes the following materials and reagents detailed in Table 1:

Reference	Reagent/Material	Description	Color/Barcode	Amount
VS-NCO312	SARS-CoV-2 (N1 + N2) reaction tube	A mix of enzymes, primers probes, buffer, dNTPs, stabilizers and endogenous internal control in stabilized format	Transparent Green or 1G foil	2 pouches of 12 tubes
VS-RB09	Rehydration Buffer tube	Solution to reconstitute the stabilized product	Transparent Orange or 11 foil	1 pouch of 24 tubes

Table 1. Reagents and materials provided in VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System with Cat. N°. VS-NCO324 (444215).



5. Reagents and equipment to be supplied by the user

The following list includes the materials and equipment that are required for use but not included in the VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System.

- Real-time PCR instrument: BD MAX™ System.
- BD MAX™ ExK™ TNA-3 (Ref:442827 or 442828)
- BD MAX™ PCR Cartridges (Ref: 437519)
- Vortex.
- Micropipettes (accurate between 2 and 1000 µL).
- Filter tips.
- Powder-free disposable gloves

6. Transport and storage conditions

- The kits can be shipped and stored at 2-40°C until the expiration date which is stated on the label.
- After opening the aluminum pouches which contain the reaction tubes can be used up to 28 days.

7. Precautions for users

- The product is intended for use by professional users only, such as laboratory or health professionals and technicians, trained in molecular biological techniques.
- For *in vitro* diagnostic use.
- Do not use expired reagents and/or materials.
- Do not use the kit if the label that seals the outer box is broken.
- Do not use reagents if the protective box is open or broken upon arrival.
- Do not use reagents if the protective pouches are open or broken upon arrival.
- Do not use reagents if desiccant is not present or broken inside reagent pouches.
- Do not remove desiccant from reagent pouches.
- Close protective pouches of reagents promptly with the zip seal after each use. Remove any excess air in the pouches prior to sealing.
- Do not use reagents if the foil has been broken or damaged.
- Do not mix reagents from different pouches and/or kits and/or lots.
- Protect reagents from humidity. Prolonged exposure to humidity may affect product performance.
- Keep components away from light.
- In cases where other PCR tests are conducted in the same general area of the laboratory, care must be taken to ensure that the VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System, BD MAX™ ExK™ TNA-3 extraction kit, any additional reagents required for testing, and the BD MAX™ System are not contaminated. Avoid microbial and ribonuclease (RNase)/deoxyribonuclease (DNase) contamination of reagents at all times. The use of sterile RNase/DNase-free disposable aerosol resistant or positive displacement pipette tips is recommended. Use a new tip for each specimen. Gloves must be changed before manipulating reagents and cartridges.



- To avoid contamination of the environment by amplicons, do not break apart the BD MAX™ PCR Cartridge after use. The seals of the BD MAX™ PCR Cartridge are designed to prevent contamination.
- Design a unidirectional workflow. It should begin in the Extraction Area and then move to the Amplification and Detection Area. Do not return samples, equipment and reagents to the area in which the previous step was performed.
- Follow Good Laboratory Practices. Wear protective clothing, use disposable gloves, goggles and mask. Do not eat, drink or smoke in the working area. Wash your hands after finishing the test.
- Samples must be treated as potentially infectious as well as all the reagents and materials that have been exposed to the samples and they must be handled according to the national safety regulations. Take necessary precautions during the collection, storage, treatment and disposal of samples.
- Regular decontamination of commonly used equipment is recommended, especially micropipettes and work surfaces.
- Consult the BD MAX™ System User's Manual for additional warnings, precautions and procedures.

8. Procedure

8.1. SAMPLE COLLECTION, STORAGE AND TRANSPORT

The VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System has been validated on nasopharyngeal/ oropharyngeal swab collected in viral transport media (VTM) Vircell S.L., Spain).

Other types of samples from nasopharyngeal/oropharyngeal swabs in VTM must be validated by the user.

Collection, storage and transport specimens should be maintained per the conditions validated by the user. Overall, respiratory samples should be collected and labelled appropriately in clean containers with or without transport media (depending on sample type) and processed as soon as possible to guarantee the quality of the test. The specimens should be transported at 2 to 8°C for up to 48 hours, following the local and national regulations for the transport of pathogen material. For long term transport (more than 48 hours), we recommend shipping at ≤ 20°C. It is recommended to use fresh specimens for the test. The samples can be stored at 2 to 8°C for up to 48 hours or frozen at -20°C or ideally at -70°C for conservation. Repeated freeze-thaw cycles should be avoided in order to prevent degradation of the sample and nucleic acids.

8.2. SAMPLE PREPARATION AND RNA EXTRACTION

Perform the sample preparation according to the recommendations in the instructions for use of extraction kit used, BD MAX™ ExK™ TNA-3. Note that some other samples may require pre-processing. Application-specific extraction preparation procedures should be developed and validated by the user.

1. Pipette between 400 and 750 µL of nasopharyngeal/ oropharyngeal swab collected in viral transport media (VTM) or in BD™ Universal Viral Transport (UVT) System media into a BD MAX™ TNA-3 Sample Buffer Tube and close the tube with a septum cap. Ensure complete mixing by vortexing the sample at high speed for 1 minute. Proceed to BD MAX™ System Operation.



8.3. PCR PROTOCOL

Note: Please, refer to the BD MAX™ System User's Manual for detailed instructions.

8.3.1. Creating PCR test program for VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System

Note: If you have already created the VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection test, you can skip step 8.3.1 and go directly to 8.3.2.

- 1) On the "Run" screen of the BD MAX™ System, select the "Test Editor" tab.
- 2) Click the "Create" button.
- 3) In the Basic Information tab, within the "Test Name" window, name your test: i.e. VIASURE SARS-CoV-2 (N1 + N2).
- 4) In the "Extraction Type" drop down menu, select "ExK TNA-3".
- 5) In the "Master Mix Format" drop down menu, choose "Type 5"
 - a. Note: Product may be used in combination with an additional VIASURE for BD MAX™ test, then select "Dual Master Mix Concentrated Lyophilized MM with Rehydration Buffer (Type 5)".
- 6) In the "Sample extraction parameters" select "User defined" and adjust sample volume to 950 µL.
- 7) In the "Ct Calculation" select "Call Ct at Threshold Crossing".
- 8) In "PCR settings" tab enter the following parameters: "Channel Settings", "Gains" and "Threshold" (Table 2).
 - a. Note: Product may be used in combination with an additional VIASURE for BD MAX™ test, PCR Settings and Test Steps should be completed for snap 2 (green) and snap 4 (blue) positions.

Channel	Alias	Gain	Threshold	Ct Min	Ct Max
475/520 (FAM)	SARS-CoV-2 N2 target	80	150	0	40
530/565 (HEX)	Endogenous IC	80	150	0	35
585/630 (ROX)	-	0	0	0	0
630/665 (Cy5)	SARS-CoV-2 N1 target	80	150	0	40
680/715 (Cy5.5)	-	0	0	0	0

Table 2. PCR settings.

Note: It is recommended to set the minimum threshold values listed above for each channel as a starting point, but the final settings must be determined by the end-user during the result interpretation in order to ensure that thresholds fall within the exponential phase of the fluorescence curves and above any background signal. The threshold value for different instruments may vary due to different signal intensities.

- 9) In "PCR settings" tab enter the following parameters "Spectral Cross Talk" (Table 3), as well.



		False Receiving Channel				
Channel		475/520	530/565	585/630	630/665	680/715
Excitation Channel	475/520	-	3.0	0.0	0.0	0.0
	530/565	1.0	-	0.0	0.0	0.0
	585/630	0.0	0.0	-	0.0	0.0
	630/665	0.0	0.0	0.0	-	0.0
	680/715	0.0	0.0	0.0	0.0	-

Table 3. Spectral cross-talk parameters.

- 10) In "Test Steps" tab, enter the PCR protocol (Table 4).

Step Name	Profile Type	Cycles	Time (s)	Temperature	Detect
Reverse transcription	Hold	1	900	45°C	-
Initial denaturation	Hold	1	120	98°C	-
Denaturation and Annealing/Extension (Data collection)	2-Temperature	45	10	95°C	-
			61.1	63°C	✓

Table 4. PCR protocol.

- 11) Click the "Save Test" button.

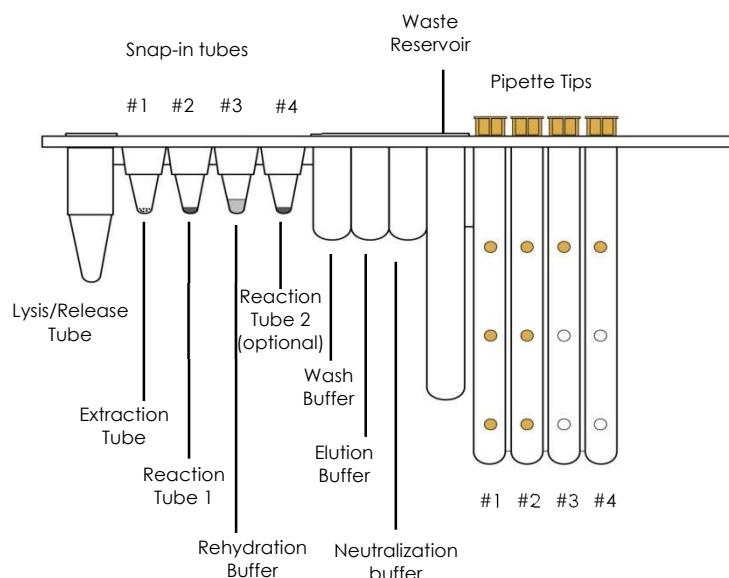
8.3.2. BD MAX™ Rack set up

- For each sample to be tested, remove one Unitized Reagent Strips from the BD MAX™ ExK TNA-3 kit. Gently tap each strip onto a hard surface to ensure that all the liquids are at the bottom of the tubes and load on the BD MAX™ System sample racks.
- Remove the required number of BD MAX™ ExK™ TNA Extraction Tubes (B4) (white foil) from their protective pouch. Snap the Extraction Tube(s) (white foil) into its corresponding positions in the TNA strip (Snap position 1, white color coding on the rack. See Figure 1). Remove excess air, and close pouch with the zip seal.
- Determine and separate the appropriate number of VIASURE SARS-CoV-2 (N1 + N2) reaction tubes (green or 1G foil) and snap into their corresponding positions in the strip (Snap position 2, green color coding on the rack. See Figure 1).
 - Remove excess air, and close aluminum pouches with the zip seal.
 - In order to carry out a correct rehydration, please make sure that the lyophilized product is in the bottom of the tube and is not adhered to the top area of the tube or to the foil seal. Gently tap each tube on a hard surface to make sure all the product is at the bottom of the tube.
 - Note: If you choose the format "Dual Master Mix Concentrated Lyophilized MM with Rehydration Buffer (Type 5)" (Section 8.3.1), determine and separate the appropriate number of additional VIASURE reaction tubes (different foil) and snap into their corresponding positions in the strip (Snap position 4, blue color coding on the rack. See Figure 1). Remove excess air, and close aluminum pouches with the zip seal.



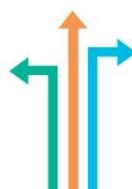
- 4) Remove the required number of Rehydration Buffer tubes (orange or 11 foil) and snap into their corresponding positions in the strip (Snap position 3, non-color coding on the rack. See Figure 1). Remove excess air, and close the pouch with the zip seal.
 - a. In order to ensure a correct transfer, please make sure that the liquid is in the bottom of the tube and is not adhered to the top area of the tube or to the foil seal. Gently tap each tube on a hard surface to make sure all the buffer is at the bottom of the tube.

Figure 1. BD MAX™ TNA Reagent Strip (TNA) from the BD MAX™ ExK TNA-3 kit.



8.3.3. BD MAX™ Instrument set up

- 1) Select the "Work List" tab on the "Run" screen of the BD MAX™ System software v4.50A or higher.
- 2) In the "Test" drop down menu, select VIASURE SARS-CoV-2 (N1 + N2) (if not already created see Section 8.3.1).
- 3) Select the appropriate kit lot number (found on the outer box of extraction kit used) from the pull down menu (optional).
- 4) Enter the Sample Buffer Tube identification number into the Sample tube window of the Worklist, either by scanning the barcode with the scanner or by manual entry.
- 5) Fill the Specimen/Patient ID and/or Accession window of the Worklist and click the "Save" button. Continue until all Sample Buffer Tubes are entered. Ensure that the specimen/patient ID and the Sample Buffer Tubes are accurately matched.
- 6) Place the prepared Sample Buffer Tube into the BD MAX™ Rack(s).
- 7) Load the rack(s) into the BD MAX™ System (Rack A is positioned on the left side of the BD MAX™ System and Rack B on the right side).
- 8) Place the required number of BD MAX™ PCR Cartridge(s) into the BD MAX™ System.
- 9) Close the BD MAX™ System door.
- 10) Click "Start Run" to begin the procedure.



8.3.4 BD MAX™ report

- 1) In main menu, click the "Results" button.
- 2) Either double click on your run in the list or press the "view button".
- 3) Click on "Print", select: "Run Details, Test Details and Plot..."
- 4) Click on "Print or Export button" on the "Run Reports" screen

9. Result interpretation

For a detailed description on how to analyze data, refer to the BD MAX™ System User's manual.

The analysis of the data is done by the BD MAX™ software according to the manufacturer's instructions. The BD MAX™ software reports Ct values and amplification curves for each detector channel of each sample tested in the following way:

- Ct value of 0 indicates that there was no Ct value calculated by the software with the specified Threshold (see Table 2). Amplification curve of the sample showing a "0" Ct value must be checked manually.
- Ct value of -1 indicates that no amplification process has occurred.
- Any other Ct value should be interpreted in correlation with the amplification curve and according to the sample interpretation guidelines outlined in Table 5.

Check Internal Control signal to verify the correct functioning of the amplification mix. In addition, check that there is no report of BD MAX™ System failure.

Results should be read and analyzed using the following table:



SARS-CoV-2 (N2 target) (475/520)	Endogenous Internal Control (530/565)	SARS-CoV-2 (N1 target) (630/665)	Interpretation
+	+/- ¹	+	SARS-CoV-2 N gene RNA Detected¹
+ ²	+/- ¹	-	SARS-CoV-2 N gene RNA Detected^{1,2}
-	+/- ¹	+ ²	SARS-CoV-2 N gene RNA Detected^{1,2}
-	+ ³	-	SARS-CoV-2 N gene RNA Not Detected³
-	- ³	-	Unresolved (UNR) Result obtained in the presence of inhibitors in the PCR reaction or when a general problem (not reported by an error code) with the sample processing and/or amplification steps occurs.³
IND	IND	IND	Indeterminate assay result (IND). Due to BD MAX™ System failure. Assay result displayed in case of an instrument failure linked to an error code.
INC	INC	INC	Incomplete assay result (INC). Due to BD MAX™ System failure. Assay result displayed in case of failure to complete run.

Table 5. Sample interpretation

+: Amplification occurred

-: No amplification occurred

1 A sample is considered positive if the Ct value obtained is less than 40. The endogenous Internal Control (IC) may or may not show an amplification signal. Sometimes, the IC detection is not necessary because a high copy number of the target can cause preferential amplification of target-specific nucleic acids.

2 If only one target site of the N gene amplifies, verify the sigmoid shape of the curve and the intensity of fluorescence. In case of a doubtful interpretation, depending on the available material, it is also recommended to:

- a) re-extract and re-test another aliquot of the same specimen (if possible, increase sample volume to 750 µl) or,
- b) obtain a new specimen and re-test.

3 In the case of SARS-CoV-2 target sites negative, IC must show an amplification signal with Ct less than 35. The Ct value could be very variable due to the Endogenous Internal Control is a human housekeeping gene that should be present in all human nucleated cells in the original sample. If there is an absence of signal or Ct value ≥ 35 of the endogenous Internal Control, the result is considered as 'Unresolved', and retesting is required.

In case of a continued ambiguous result, it is recommended to review the instructions for use, the extraction process used by the user; to verify the correct performance of each RT-qPCR steps and review the parameters; and to check the sigmoid shape of the curve and the intensity of fluorescence.

The results of the test should be evaluated by a health care professional in the context of medical history, clinical symptoms and other diagnostic tests.



10. Limitations of the test

- The results of the test should be evaluated by a health care professional in the context of medical history, clinical symptoms and other diagnostic tests.
- Although this assay can be used with other types of samples it has been validated with nasopharyngeal/oropharyngeal swab collected in VTM.
- For good test performance, the lyophilized product should be at the bottom of the tube and not adhered to the top area of the tube or the foil seal. Gently tap each tube on a hard surface to make sure all the product is at the bottom of the tube.
- An appearance of the reaction mixture in stabilized format, normally found at the bottom of the tube, different from the usual one (without conical shape, inhomogeneous, smaller/larger in size and/or color different from whitish) does not alter the functionality of the test.
- The quality of the test depends on the quality of the sample; proper extracted nucleic acid from respiratory samples must be extracted.
- This test is a qualitative test and does not provide quantitative values or indicate the number of organisms present.
- Extremely low levels of target below the limit of detection might be detected, but results may not be reproducible.
- There is a possibility of false positive results due to cross-contamination by SARS-CoV-2, either samples containing high concentrations of target RNA or contamination due to PCR products from previous reactions.
- The specific primer and probe combinations for detection of conserved regions of N gene used in VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System have been designed based on the US CDC assay for specific detection of SARS-CoV-2 by amplifying two unique regions of the N gene. They do not show significant combined homologies with the human genome, human microflora, SARS-CoV or other coronaviruses, which might result in predictable false positive.
- False Negative results may arise from several factors and their combinations, including:
 - Improper specimens' collection, transport, storage, and/or handling methods.
 - Improper processing procedures (including RNA extraction).
 - Degradation of the viral RNA during sample shipping/storage and/or processing.
 - Mutations or polymorphisms in primer or probe binding regions may affect detection of new or unknown SARS-CoV-2 variants.
 - A viral load in the specimen below the limit of detection for the assay.
 - The presence of RT-qPCR inhibitors or other types of interfering substances. The impacts of vaccines, antiviral therapeutics, antibiotics, chemotherapeutics or immunosuppressant drugs used to prevent COVID-19 or used during the treatment of the infection have not been evaluated.
 - Failure to follow instructions for use and the assay procedure.
- A single-target site amplification or even random positive results is suggestive of slightly different amplification yield of the target site of the N gene. Samples with low viral load might result in N single target amplification. In case of a doubt, it is recommended referring to a reference laboratory for further testing.



- Some samples may fail to exhibit RNase P amplification curves due to low human cell numbers in the original clinical sample. A negative IC signal does not preclude the presence of SARS-CoV-2 RNA in a clinical specimen.
- A positive test result does not necessarily indicate the presence of viable viruses and does not imply that these viruses are infectious or are the causative agents for clinical symptoms. However, a positive result is indicative of the presence of targets viral sequences (N genes).
- Negative results do not preclude SARS-CoV-2 infection and should not be used as the sole basis for treatment or other patient management decisions. Optimum specimen types and timing for peak viral levels during infections caused by SARS-CoV-2 have not been determined. The collection of multiple specimens (types and time points) from the same patient may be necessary to detect the virus.
- If diagnostic tests for other respiratory illnesses are negative and the patient's clinical presentation and epidemiological information suggest that SARS-CoV-2 infection is possible, then a false negative result should be considered, and a re-testing of the patient should be discussed.
- In the case of obtaining Unresolved, Indeterminate or Incomplete results using VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System retesting will be required. Unresolved results may be due to the presence of inhibitors in the sample or an incorrect rehydration of lyophilized reaction mix tube. If there is an instrument failure, Indeterminate or Incomplete results will be obtained.

11. Quality control

VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System contains an endogenous internal control (IC) in each reaction tube which confirms the correct performance of the technique.

12. Performance characteristics

12.1. Clinical sensitivity and specificity

The clinical performance of VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System was tested using 254 respiratory samples (nasopharyngeal swabs in Vircell Transport medium) from patients with clinical suspicion of COVID-19 disease or other similar respiratory diseases. The retrospective-comparative analysis was performed with VIASURE SARS-CoV-2 (N1 + N2) Real time PCR Kit for BD MAX™ System and these results were compared with those obtained with the clinical diagnosis performed with Simplexa™ COVID-19 Direct assay with discrepant analysis performed with the Charité protocol.

	Alternative RT-PCR assays		
		+	-
+	63	2*	65
-	0	189	189
Total	63	191	254

Table 6. Comparative results for SARS-CoV-2.

*Initial diagnose of one of the two samples was invalid and reported to the patient as positive for prevention and quarantine period.



VIASURE SARS-CoV-2 (N1 + N2) Real time PCR Kit for BD MAX™ System detected two positive samples that were not detected using Simplexa™ COVID-19 Direct assay and the Charité protocol.

The Positive Percent Agreement (PPA) and the Negative Percent Agreement (NPA) for VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR detection Kit for BD MAX™ System are >99% and 98%, respectively.

Results show high agreement to detect SARS-CoV-2 using VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System.

12.2. Analytical sensitivity

VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System has a detection limit of ≥ 5 genome copies per reaction with a positive rate of $\geq 95\%$.

Figure 2. Dilution series of SARS-CoV-2 (N1 + N2) (9.9×10^4 - 9.9×10^0 and 5.0×10^0 genome copies per reaction) template run on the BD MAX™ System (475/520 (FAM) channel).

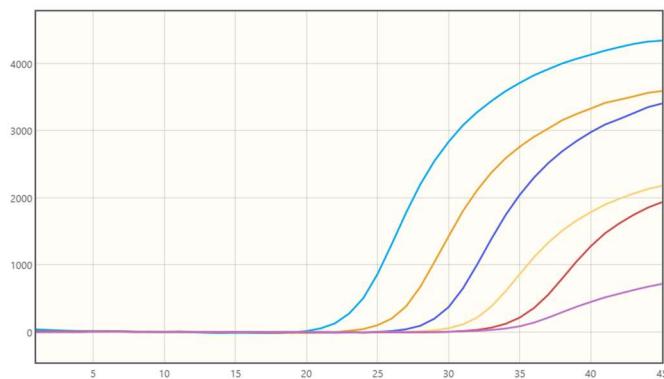
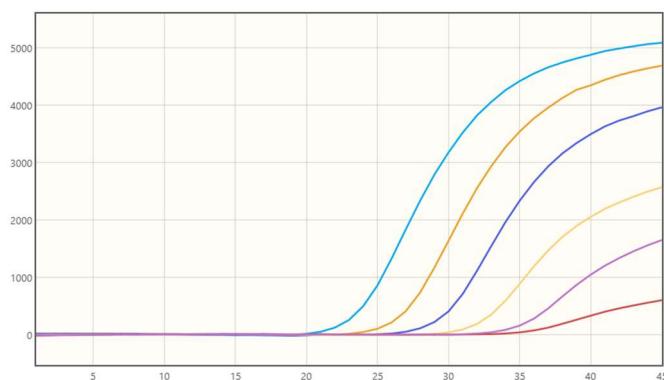


Figure 3. Dilution series of SARS-CoV-2 (N1 + N2) (9.9×10^4 - 9.9×10^0 and 5.0×10^0 genome copies per reaction) template run on the BD MAX™ System (630/665 (Cy5) channel).



12.3. Analytical specificity

The specificity of the SARS-CoV-2 (N1 + N2) assay was confirmed by testing a panel consisting of different microorganisms representing the most common respiratory pathogens. No cross-reactivity was detected between any of the following microorganisms tested:



Cross-reactivity testing				
Human Adenovirus types 1-5, 8, 15, 31, 40 and 41	-	Influenza A/New Caledonia/20/99(H1N1) virus	-	<i>Legionella longbeachae</i>
Human Bocavirus	-	Influenza A/California/7/2009(H1N1)pdm09 virus	-	<i>Legionella micdadei</i>
<i>Bordetella bronchiseptica</i>	-	Influenza A/Michigan/45/2015 (H1N1)pdm09 virus	-	<i>Legionella pneumophila</i>
<i>Bordetella holmesii</i>	-	Influenza A/Singapore/GP1908/2015, IVR-180 (H1N1)pdm09 virus	-	Human metapneumovirus A and B
<i>Bordetella parapertussis</i>	-	Influenza A/Victoria/210/2009 (H3N2) virus	-	<i>Moraxella catarrhalis</i>
<i>Bordetella pertussis</i>	-	Influenza A/Thüringen/5/2017 (H3N2) virus	-	<i>Mycoplasma pneumoniae</i>
<i>Chlamydia caviae</i>	-	Influenza A/Switzerland/9715293/2013 (H3N2) virus	-	<i>Mycobacterium tuberculosis</i> not rifampin resistant
<i>Chlamydia psittaci</i> genotype A and C	-	Influenza A/Hong Kong/4801/2014, NYMC X-263B (H3N2) virus	-	Human parainfluenza 1, 2, 3 and 4 viruses
<i>Chlamydophila pneumoniae</i> CM-1	-	Influenza A/ South Australia/55/2014, IVR-175 (H3N2) virus	-	<i>Pneumocytis jirovecii</i> Type A1 and g885652
Human coronavirus 229E, OC43, NL63 and HKU1	-	Influenza A/DE-SH/Reiherente/AR8444/ 2016 (H5N8) virus	-	Human rhinovirus type C
MERS Coronavirus	-	Influenza A/Anhui/1/2013 (H7N9) virus	-	<i>Staphylococcus aureus</i> subsp. <i>aureus</i>
SARS Coronavirus Strain Frankfurt 1	-	Influenza B/Brisbane/60/2008 virus	-	<i>Staphylococcus epidermidis</i>
Enterovirus 68 and 71	-	Influenza B/Florida/04/06 virus	-	<i>Streptococcus pneumoniae</i> Z022
Enterovirus Echovirus 11 and 30	-	Influenza B/Phuket/3073/2013 virus	-	<i>Streptococcus pyogenes</i>
Enterovirus Coxsackievirus A24, A9 and B3	-	<i>Legionella bozemanii</i>	-	<i>Streptococcus salivarius</i>
<i>Haemophilus influenzae</i> MinnA	-	<i>Legionella dumoffii</i>	-	Respiratory syncytial virus (RSV) A and B

Table 7. Reference pathogenic microorganisms used in this study.

12.4. Analytical reactivity

The reactivity of VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System was evaluated against RNA from Human 2019-nCoV strain BetaCoV/Germany/BavPat1/2020 p.1, Human 2019-nCoV strain 2019-nCoV/Italy-INMI1, SARS-CoV-2 strain 2019nCoV/USA-WA1/2020, synthetic RNA controls for two variants of the SARS-CoV-2 virus: MT007544.1 (SARS-CoV2 isolate Australia/VIC01/2020) and MN908947.3 (SARS-CoV-2 isolate Wuhan-Hu-1), showing positive result.



HRVATSKI

1. Namjena

VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System je automatski RT-PCR test u stvarnom vremenu osmišljen za kvalitativnu detekciju RNK iz SARS-CoV-2 u uzorcima iz dišnog sustava od pojedinaca sa sumnjom na COVID-19 od strane njihovih liječnika. Ovaj test je namijenjen za uporabu kao pomagalo u identifikaciji prisutnosti SARS-CoV-2 virusne RNK. Test koristi BD MAX™ System za automatiziranu ekstrakciju RNK, a zatim lančanu reakciju polimeraze RT-PCR u stvarnom vremenu s priloženim reagensima u kombinaciji s univerzalnim reagensima i jednokratnim materijalom za BD MAX™ System. RNK se ekstrahira iz uzorka iz dišnog sustava, amplificirana uporabom tehnike RT-PCR te detektirana uporabom fluorescentne sonde za boju reporteta specifične za SARS-CoV-2.

2. Sažetak i objašnjenje

Koronavirusi su grupa ovijenih, nesegmentiranih, pozitivno usmjerenih RNK virusa koji spadaju u obitelj Coronaviridae [1,2]. Poznato je šest vrsta koronavirusa koji uzrokuju bolesti u ljudi [2]. Četiri virusa (229E, OC43, NL63 i HKU1) uzrokuju uobičajene simptome prehlade, a preostala dva (koronavirus teškog akutnog respiratornog sindroma (SARS-CoV) i koronavirus respiratornog sindroma Bliskog istoka (MERS-CoV)) su zoonotični i izazivaju teže komplikacije [2]. SARS-CoV i MERS-CoV prouzročili su preko 10.000 kumulativnih slučajeva u protekla dva desetljeća, sa stopama smrtnosti od 34% MERS-CoV i 10% SARS-CoV [1,3].

U prosincu 2019., nekoliko osoba koje su radile i živjele oko tržnice morskih plodova i životinja Huanan u Wuhanu, kineskoj pokrajini Hubei, oboljele su od upale pluća nepoznatog uzroka [2,4]. Analiza dubokog sekvenciranja respiratornih uzorka ukazala je na novi koronavirus, koji je prvo dobio ime novi koronavirus 2019 (2019-nCoV), a zatim SARS-CoV-2 [5].

Potvrđen je prijenos SARS-CoV-2 s čovjeka na čovjeka, čak i tijekom inkubacijskog razdoblja bez simptoma, a virus uzrokuje teške respiratorne bolesti poput onih izazvanih virusom SARS-CoV [1,6,7,8]. Iako je upala pluća najčešća povezana bolest, u nekoliko bolesnika razvila se teška upala pluća, plućni edem, akutni respiratori distres sindrom ili zakazivanje više organa i smrt [1,4]. Centri za kontrolu i prevenciju bolesti (Centers of Disease Control and Prevention, CDC) smatraju da se simptomi SARS-CoV-2 mogu pojaviti od 2 do 14 dana nakon izlaganja, pritom su najčešći zimica, kašalj, umor, anoreksija, mijalgija i dispneja [1,4,6,9]. Manje česti simptomi su grlobolja, začepljeno nos, glavobolja, proljev, mučnina i povraćanje [1,4]. Prijavljeni su također i gubitak mirisa (anozmija) ili gubitak okusa (ageuzija) prethode nastupu respiratornih simptoma [9]. Stariji odrasli i osobe koje imaju teška postojeća medicinska stanja poput bolesti srca ili pluća ili dijabetesa pod većim su rizikom od razvoja ozbiljnijih komplikacija bolesti COVID-19 [10].

Dijagnosticiranje SARS-CoV-2 provodi se ranom detekcijom konvencionalnih uzroka upale pluća i detektira se sekvenciranjem sljedeće generacije ili metodama lančane reakcije polimeraze reverznom transkripcijom (RT-PCR) u stvarnom vremenu [1,11]. Trenutno je dostupno nekoliko testova koji detektiraju SARS-CoV-2 poput Kina CDC (ciljanje gena, ORF1ab i N), Charité – Njemačka (ciljanje gena, RdRP ili E) ili SAD CDC (dva cilja u N genu) [12].



Centar za kontrolu bolesti preporučuje uzorke iz gornjeg dijela dišnog trakta (nazofaringealni (NP) i orofaringealni (OP) brisevi, bris srednje turbine nosa, nazalni bris, uzorci nazofaringealnog ispirka/aspirata ili nazalnog ispirka/aspirata (NW) prikupljeni uglavnom od strane zdravstvenog radnika) i/ili uzorci donjeg dijela dišnog sustava (ispitjuvacki, endotrahealni aspirat ili bronhoalveolarna lavaža u bolesnika s težom bolešću dišnih putova) za identifikaciju SARS-CoV-2 [11]. Pored toga, mogu se prikupiti i drugi klinički uzorci poput krvi, mokraće i stolice za nadzor prisutnosti virusa [11,12].

3. Načelo postupka

Komplet za detekciju VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System osmišljen je za identifikaciju SARS-CoV-2 u uzorcima iz dišnog sustava. Detekcija se obavlja u obliku lančane reakcije polimeraze reverznom transkripcijom (RT-PCR) u stvarnom vremenu koja se sastoji od jednog koraka, gdje se reverzna transkripcija i potom amplifikacija specifične ciljne sekvene vrši u istoj reakcijskoj epruveti. Nakon izolacije ciljne RNK, vrši se njena transkripcija čime se dobiva komplementarna DNK zahvaljujući reverznoj transkriptazi, nakon koje slijedi amplifikacija dvije konzervirane regije gena N (N1+N2) primjenom specifičnih početnica i fluorescentno obojane sonde.

VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System temelji se na aktivnosti 5' egzonukleaze DNK polimeraze. Tijekom amplifikacije DNK ovaj enzim rascjepljuje sondu vezanu za sekvenu komplementarne DNK, čime se razdvaja supresorsku boju od indikacijskog gena. Ova reakcija dovodi do povećanja fluorescencijskog signala srazmernog količini ciljnog predloška. Ta fluorescencija mjeri se na BD MAX™ sustavu.

Komplet za detekciju VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System sadrži u svakoj epruveti sve komponente potrebne za test PCR (specifične primere/sonde, dNTPS, pufer, polimerazu, reverznu transkriptazu) u stabiliziranom formatu, kao i endogenu unutarnju kontrolu za praćenje procesa ekstrakcije i/ili inhibicije aktivnosti polimeraze. Test koristi humani domaćinski gen kao endogenu internu kontrolu (IC) (humani RNase P gen). Ljudski domaćinski geni uključeni su u osnovno održavanje stanice te se stoga očekuje da će biti prisutni u svim ljudskim stanicama s jezgrom te održavati relativno stalne razine ekspresije. Cilj N2 je pojačan i detektiran u kanalu 475/520, cilj N1 u kanalu 630/665 te endogenoj internoj kontroli (IC) u kanalu 530/565.

4. Reagensi koji se isporučuju

Komplet za detekciju VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System uključuje sljedeće materijale i reagense detaljno opisane u Tablici 1:



Referenca	Reagens/materijal	Opis	Boja/crtični kod	Količina
VS-NCO312	SARS-CoV-2 (N1 + N2) reaction tube	Smjesa enzima, sondi za početnice, pufera, dNTP-ova, stabilizatora i endogene unutarnje kontrole u stabiliziranom obliku	Prozirno Zelena ili 1G folija	2 vrećice s 12 epruveta
VS-RB09	Rehydration Buffer tube	Otopina za rekonstituciju stabiliziranog proizvoda	Prozirno Narančasta ili 11 folija	1 vrećica s 24 epruvete

Tablica 1. Reagensi i materijali osigurani u kompletu za detekciju VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System with Cat. N°. VS-NCO324 (444215).

5. Reagensi i oprema koju mora pribaviti korisnik

Sljedeći popis uključuje materijale i opremu koja je neophodna za korištenje, ali nije dio kompleta za detekciju VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System.

- Instrument za lančanu reakciju polimeraze (PCR) u stvarnom vremenu: BD MAX™ System.
- BD MAX™ ExK™ TNA-3 (Ref:442827 ili 442828)
- BD MAX™ PCR Cartridges (Ref: 437519)
- Vortex.
- Mikropipete (precizne u rasponu od 2 do 1000 µl).
- Nastavci za filter.
- Nenapršene jednokratne rukavice.

6. Uvjeti za transport i pohranu

- Kompleti se mogu transportirati i čuvati na temperaturi od 2-40°C do isteka roka valjanosti navedenog na naljepnicama.
- Nakon otvaranja aluminijskih vrećica koje sadrže reakcijske epruvete mogu se iskoristiti u roku od 28 dana.

7. Mjere opreza za korisnike

- Proizvod je namijenjen isključivo uporabi od strane profesionalnih korisnika, kao što su laboratorijski ili zdravstveni djelatnici i tehničari, koji su prošli edukaciju o tehnikama molekularne biologije.
- Samo za *in vitro* dijagnostičku upotrebu.
- Nemojte koristiti reagense i/ili materijale kojima je istekao rok valjanosti.
- Nemojte koristiti komplet ako je potrgana naljepnica kojom je zatvorena vanjska kutija.
- Nemojte koristiti reagense ako je zaštitna kutija otvorena ili oštećena po prispjeću.
- Nemojte koristiti reagense ako su zaštitne vrećice otvorene ili oštećene po prispjeću.
- Nemojte koristiti reagense ako desikant u vrećicama s reagensima nije prisutan ili je oštećen.
- Nemojte vaditi desikant iz vrećica s reagensima.
- Nakon svake upotrebe odmah zatvorite zaštitne vrećice s reagensima pomoću patentnog zatvarača. Prije zatvaranja istisnite višak zraka iz vrećica.
- Nemojte koristiti reagense ako je folija potrgana ili oštećena.



- Nemojte miješati reagense iz različitih vrećica, kompleta i/ili serija.
- Zaštite reagense od vlage. Duže izlaganje vlazi može utjecati na učinak proizvoda.
- Čuvajte komponente dalje od svjetlosti.
- U slučajevima gdje se drugi testovi PCR-om obavljaju u istom općem prostoru laboratorija potreban je oprez kako ne bi došlo do kontaminacije kompleta VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System, BD MAX™ ExK™ TNA-3 extraction kit, svih dodatnih reagensa potrebnih za testiranje i BD MAX™ System. Svakako izbjegavajte kontaminaciju reagensa s mikrobima i ribonukleazama (Rnase)/deoksiribonukleazama (Dnase). Preporučuje se uporaba jednokratnih, sterilnih vrhova volumetrijskih pipeta ili otpornih na aerosol, bez Rnase/Dnase. Koristite novi vrh sa svakim uzorkom. Rukavice se moraju promijeniti prije rukovanja reagensima i patronama.
- Radi sprječavanja kontaminacije okruženja amplikonima, nemojte odvajati BD MAX™ PCR Cartridge nakon uporabe. Brtve na BD MAX™ PCR Cartridge dizajnirane su za sprječavanje kontaminacije.
- Koristite jednosmjeran radni proces. Treba započeti u području za ekstrakciju, a zatim preći u područje za amplifikaciju i detekciju. nemojte vraćati uzorke, opremu i reagense u područje u kojem je obavljen prethodni korak.
- Pridržavajte se dobre laboratorijske prakse. Nosite zaštitnu odjeću, jednokratne rukavice, zaštitne naočale i masku. Nemojte jesti, piti niti pušiti u radnom prostoru. Operite ruke nakon što završite test.
- Uzorci se moraju smatrati potencijalno zaraznim, a isto vrijedi za reagense i materijale koji su bili izloženi uzorcima i mora se rukovati u skladu s nacionalnim sigurnosnim propisima. Poduzmite neophodne mjere opreza prilikom prikupljanja, pohrane, tretiranja i odlaganja uzorka u otpad.
- Preporučuje se redovita dekontaminacija opreme koja se često koristi, naročito mikropipeta i radnih površina.
- Posavjetujte se s Korisničkim priručnikom za BD MAX™ sustav u vezi s dodatnim upozorenjima, mjerama opreza i postupcima.

8. Postupak

8.1. PRIKUPLJANJE, POHRANA I TRANSPORT UZORAKA

Komplet za detekciju VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System validiran je na nazofaringealnom/orofaringelnom brisu prikupljenom u virusnom transportnom mediju (VTM) Vircell S.L., Španjolska).

Drukčije vrste uzoraka iz nazofaringealnih/orofaringealnih brisova u VTM-u mora validirati korisnik.

Prikupljanje, pohrana i transport uzorka treba obavljati u uvjetima koje je validirao korisnik. U globalu, respiratorne uzorke treba prikupiti i označiti pravilno u čiste spremnike sa ili bez transportnog medija (ovisno o vrsti uzorka) i obraditi što je moguće prije u cilju osiguranja kvalitete testa. Uzorci se trebaju transportirati na temperaturi od 2 °C do 8 °C tijekom 48 sati u skladu s lokalnim i nacionalnim propisima za transport patogenih materijala. Za dugotrajni transport (duži od 48 sata) preporučujemo otpremanje na temperaturi od ≤-20 °C. Preporučuje se upotreba svježih uzorka za test. Uzorci se mogu čuvati na temperaturi od 2 °C do 8 °C tijekom 48 sati ili držati smrznuti na temperaturi od -20°C ili idealno na -70°C za pohranu. Treba izbjegavati ponavljanje ciklusa smrzavanja i odmrzavanja kako bi se spriječilo propadanje uzorka i nukleinskih kiselina.



8.2. PRIPREMA UZORAKA I EKSTRAKCIJA RNK

Prikupljanje uzorka obavite u skladu s preporukama iz uputa za upotrebu kompleta za ekstrakciju koji koristite, BD MAX™ ExK™ TNA-3. Moguće je da će neki drugi uzorci zahtijevati predobradu. Korisnik treba razviti i validirati postupke pripreme za ekstrakciju specifične za predmetnu primjenu.

1. Pipetom prenesite između 400 µl i 750 µl nazofaringealnog/orofaringealnog brisa prikupljenog u virusnom transportnom mediju (VTM) ili u BD™ Universal Viral Transport (UVT) System medija u BD MAX™ TNA-3 Sample Buffer Tube epruvetu za uzorak s puferom te je zatvorite čepom s membranom. Izmiješajte uzorak vrtloženjem pri visokoj brzini tijekom 1 minute. Pređite na rad sa BD MAX™ sustavom.

8.3. PROTOKOL ZA PCR

Napomena: Detaljne upute potražite u Korisničkom priručniku za BD MAX™ sustav.

8.3.1. Kreiranje programa za testiranje VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System

Napomena: Ako ste već kreirali VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection test, možete preskočiti korak 8.3.1 i preći izravno na 8.3.2.

- 1) Na zaslonu „Run“ (Pokreni) na BD MAX™ System odaberite karticu „Test Editor“ (Uređivač testa).
- 2) Kliknite na gumb „Create“ (Kreiraj).
- 3) Na kartici Basic Information (Osnovne informacije) unuta prozora "Test Name" (Naziv testa), imenujte svoj test: tj. VIASURE SARS-CoV-2 (N1 + N2).
- 4) S padajućeg izbornika „Extraction Type“ (Vrsta ekstrakcije) odaberite „ExK TNA-3“.
- 5) S padajućeg izbornika „Master Mix Format“ (Oblik glavne smjese) odaberite „Type 5“ (Tip 5)
 - a. Napomena: Proizvod se može koristiti u kombinaciji s dodatnim kompletom VIASURE za BD MAX™ test, a u tom slučaju odaberite opciju „Dual Master Mix Concentrated Lyophilized MM with Rehydration Buffer (Type 5)“.
- 6) U „Sample extraction parameters“ (Parametri ekstrakcije uzorka) odaberite „User defined“ (Korisnički definirani) i prilagodite volumen uzorka na 950 µl.
- 7) U „Ct Calculation“ (Ct izračun) odaberite „Call Ct at Threshold Crossing“ (Izračunaj Ct pri prelasku granice).
- 8) Na kartici „PCR settings“ (Postavke za PCR) unesite sljedeće parametre: „Channel Settings“ (Postavke kanala), „Gains“ (Dobit) i „Threshold“ (Granica) (Tablica 2).
 - a. Napomena: Proizvod se može koristiti u kombinaciji s dodatnim kompletom VIASURE za BD MAX test, a u tom slučaju „PCR Settings“ (Postavke za PCR) i „Test Steps“ (Koraci testa) treba popuniti za položaje 2 (zeleni) i 4 (plavi).



Channel (Kanal)	Alias (Drugi nazivi)	Gain (Dobit)	Threshold (Prag)	Ct Min (Ct Min)	Ct Max (Ct Maks)
475/520 (FAM)	SARS-CoV-2 N2 cilj	80	150	0	40
530/565 (HEX)	Endogeni IC	80	150	0	35
585/630 (ROX)	-	0	0	0	0
630/665 (Cy5)	SARS-CoV-2 N1 cilj	80	150	0	40
680/715 (Cy5.5)	-	0	0	0	0

Tablica 2. Postavke za PCR.

Napomena: Preporučuje se postavljanje gore navedenih minimalnih graničnih vrijednosti za svaki kanal kao početnu točku, ali završne postavke mora odrediti krajnji korisnik tijekom tumačenja rezultata kako bi se osiguralo da granice spadaju u eksponencijalnu fazu krivulja fluorescencije i iznad bilo kojeg pozadinskog signala. Granična vrijednost različitih instrumenata može se razlikovati zbog različitih intenziteta signala.

- 9) Na kartici „PCR settings“ (Postavke za PCR) unesite i sljedeće parametre: „Spectral Cross Talk“ (Tablica 3)

		False Receiving Channel (Kanal s lažnim rezultatima)					
		Channel (Kanal)	475/520	530/565	585/630	630/665	680/715
Excitation Channel (Kanal za pobuđivanje)	475/520	-	3,0	0,0	0,0	0,0	
	530/565	1,0	-	0,0	0,0	0,0	
	585/630	0,0	0,0	-	0,0	0,0	
	630/665	0,0	0,0	0,0	-	0,0	
	680/715	0,0	0,0	0,0	0,0	-	

Tablica 3. Parametri spektralnog preklapanja signala

- 10) Na kartici „Test Steps“ (Koraci testa) unesite protokol za PCR (Tablica 4).

Step Name (Naziv koraka)	Profile Type (Vrsta profila)	Cycles (Ciklusi)	Time (s) (Vrijeme(Vremena))	Temperature (Temperatura)	Detect (Detekcija)
Reverse transcription (Reverzna transkripcija)	Čekanje	1	900	45 °C	-
Initial denaturation (Početna denaturacija)	Čekanje	1	120	98 °C	-
Denaturation and Annealing/Extension (Data collection) (Denaturacija i sparivanje/produljenje (prikljicanje podataka))	2-temperatura	45	10	95 °C	-
			61,1	63 °C	✓

Tablica 4. Protokol za PCR.

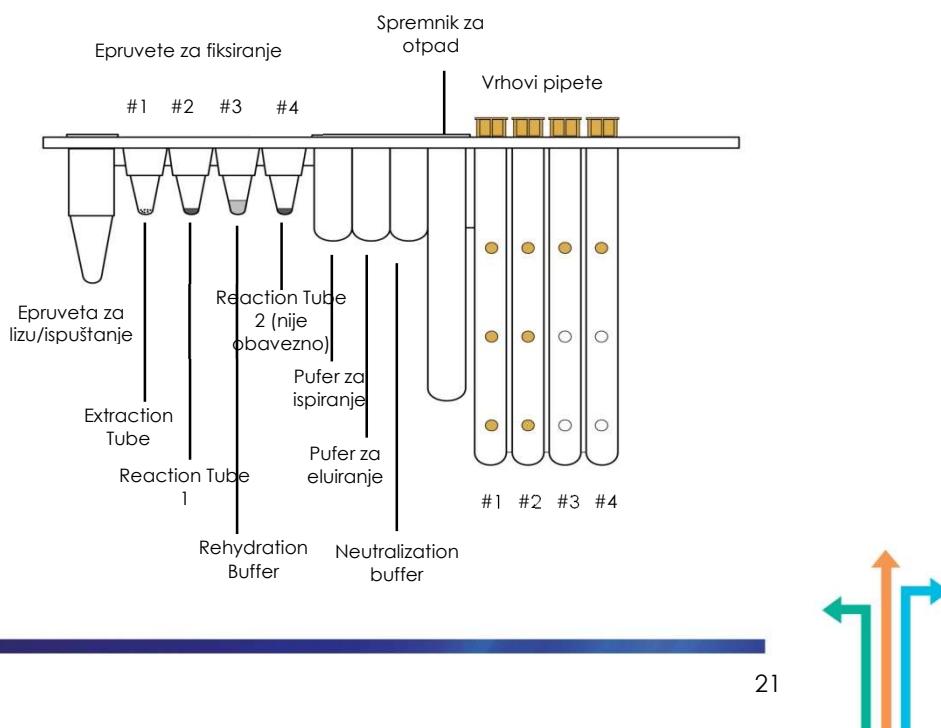
- 11) Kliknite na gumb „Save Test“ (Spremi test).

8.3.2. Postavljanje BD MAX™ stakla



- 1) Za svaki uzorak koji treba testirati izvadite pojedinačne trake s reagensima u bloku iz kompleta BD MAX™ ExK TNA-3 kit. Lagano udarite svaku traku na čvrstu površinu kako biste osigurali da se sve tekućine nalaze na dnu epruveta i stavite ih na stalke za uzorke BD MAX™ sustava.
- 2) Izvadite potrebnii broj BD MAX™ ExK™ TNA Extraction Tubes (B4) (bijela folija) iz njihove zaštitne vrećice. Postavite epruvetu/e za ekstrakciju (bijela folija) u njihove odgovarajuće položaje u TNA traku (položaj 1, bijela boja kodiranje na stalku. Pogledajte Sliku 1). Istisnite višak zraka i zatvorite vrećicu patentnom zatvaračem.
- 3) Odredite i razdvojite odgovarajući broj VIASURE SARS-CoV-2 (N1 + N2) reaction tubes (zeleni folija ili 1G folija) i postavite ih u njihove odgovarajuće položaje na traci (Snap položaj 2, označen zelenom bojom na stalku). Pogledajte Sliku 1).
 - a. Istisnite višak zraka i zatvorite aluminijске vrećice patentnim zatvaračem.
 - b. U cilju obavljanja pravilnog postupka rehidracije pazite da se liofilizirani proizvod nalazi na dnu epruvete te da ne prianja za gornji dio epruvete ili za zatvarač od folije. Lagano pritisnite svaku epruvetu na tvrdoj površini kako biste se uvjerili da je sav proizvod na dnu epruvete.
 - i. Napomena: Ako odaberete oblik „Dual Master Mix Concentrated Lyophilized MM with rehydration Buffer (Type 5)“ (Dvostruka glavna smjesa koncentriran liofiliziran MM s rehidracijskim puferom - Tip 5) (odjeljak 8.3.1), odredite i razdvojite odgovarajući broj dodatnih reakcijskih epruveta VIASURE reaction tube (drugačija folija) i postavite ih u njihove odgovarajuće položaje na traci (položaj 4, označen plavom bojom na stalku. Pogledajte Sliku 1). Istisnite višak zraka i zatvorite aluminijске vrećice patentnim zatvaračem.
- 4) Izvadite odgovarajući broj epruveta Rehydration Buffer tubes (narančasta folija ili 11 folija) i postavite ih u njihove odgovarajuće položaje na traci (položaj 3, nije obojen na stalku. Pogledajte Sliku 1). Istisnite višak zraka i zatvorite vrećicu patentnim zatvaračem.
 - a. U cilju osiguravanja pravilnog prijenosa pazite da se tekućina nalazi na dnu epruvete te da ne prianja za gornji dio epruvete ili za zatvarač od folije. Lagano pritisnite svaku epruvetu na tvrdoj površini kako biste se uvjerili da je sav proizvod na dnu epruvete.

Slika 1. BD MAX™ TNA traka s reagensima (TNA) iz kompleta BD MAX™ ExK TNA-3 kit.



8.3.3. Postavljanje BD MAX™ instrumenta

- 1) Odaberite karticu „Work List“ (Radni popis) na zaslonu „Run“ (Pokreni) na BD MAX™ sustavu s verzijom softvera v4.50A ili novijom.
- 2) S padajućeg izbornika „Test“ odaberite VIASURE SARS-CoV-2 (N1 + N2) (ako test nije već kreiran, pogledajte odjeljak 8.3.1).
- 3) Odaberite odgovarajući broj serije kompleta (naveden na vanjskoj kutiji kompleta za ekstrakciju kojeg koristite) s padajućeg izbornika (neobavezno).
- 4) Unesite identifikacijski broj epruvete za uzorak s puferom u prozor „Sample tube“ (Epruveta s uzorkom) na kartici „Worklist“ (Radni popis) skeniranjem crtičnog koda pomoću sekenera ili ručnim unošenjem broja.
- 5) Popunite polje „Specimen/Patient ID“ (ID uzorka/bolesnika) i/ili prozor „Accession“ (Pristup) na kartici „Worklist“ (Radni popis) i kliknite na gumb „Save“ (Spremi). Nastavite s postupkom dok ne unesete sve epruvete za uzorak s puferom. Pazite da se ID uzorka/bolesnika i epruvete za uzorak s puferom podudaraju.
- 6) Stavite pripremljenu epruvetu za uzorak s puferom na BD MAX™ stalak/e.
- 7) Stavite stalak/e u BD MAX™ System (stalak A se nalazi lijevo u odnosu na BD MAX™ System, dok se stalak B nalazi na desnoj strani).
- 8) Stavite potrebnii broj uložaka BD MAX™ PCR Cartridge(s) u BD MAX™ System.
- 9) Zatvorite vrata BD MAX™ System.
- 10) Kliknite „Start Run“ (Pokreni postupak) da započnete postupak.

8.3.4 BD MAX™ izvješće

- 1) U glavnom izborniku kliknite na gumb „Results“ (Rezultati).
- 2) Kliknite dvaput na svoj postupak na popisu ili pritisnite gumb „view“ (pričaz).
- 3) Kliknite na „Print“ (Ispiši) i odaberite: “Run Details, Test Details and Plot...” (Podaci o postupku, podaci o testu i grafikon...“)
- 4) Kliknite na gumb „Print or Export“ (ispisi ili izvezi) na zaslonu „Run Reports“ (Izvješća o postupku)

9. Tumačenje rezultata

Detaljne upute za analiziranje podataka potražite u Korisničkom priručniku za BD MAX™ sustav.

Analizu podataka obavlja BD MAX™ softver prema uputama proizvođača. BD MAX™ softver kreira izvješće o Ct vrijednostima i amplifikacijskim krivuljama za svaki kanal detekcije svakog uzorka testiranog na sljedeći način:

- Ct vrijednost 0 znači da softver nije izračunao Ct vrijednost s navedenom granicom (pogledajte Tablicu 2). Amplifikacijska krivulja uzorka koja pokazuje „0“ za Ct vrijednost mora se provjeriti ručno.
- Ct vrijednost -1 znači da nije došlo do amplifikacijskog procesa.



- Svaku drugu Ct vrijednost treba tumačiti u vezi s amplifikacijskom krivuljom i prema smjernicama za tumačenje uzorka navedenim u Tablici 5.

Provjerite unutarnji kontrolni signal da potvrdite funkcionira li amplifikacijska smjesa ispravno. Pored toga, provjerite da nema izvješća o kvaru BD MAX™ sustava.

Pomoću sljedeće tablice očitajte i analizirajte rezultate:

SARS-CoV-2 (N2 cilj) (475/520)	Endogena unutarnja kontrola (530/565)	SARS-CoV-2 (N1 cilj) (630/665)	Tumačenje
+	+/- ¹	+	SARS-CoV-2 N genska RNK detektirana ¹
+ ²	+/- ¹	-	SARS-CoV-2 N genska RNK detektirana ^{1,2}
-	+/- ¹	+ ²	SARS-CoV-2 N genska RNK detektirana ^{1,2}
-	+ ³	-	SARS-CoV-2 N genska RNK nije otkrivena³
-	- ³	-	Neriješeni (UNR) Rrezultat dobiven u prisutnosti inhibitora reakcije PCR-a ili kada dođe do nekog općeg problema (koji nije pokriven šifrom o pogrešci) pri obradi uzorka i/ili amplifikaciji.³
IND	IND	IND	Rezultat testa nije moguće utvrditi (IND). Zbog kvara BD MAX™ System. Rezultat testa prikazan u slučaju kvara instrumenta povezanog sa šifrom o pogrešci.
INC	INC	INC	Nepotpun rezultat testa (INC). Zbog kvara BD MAX™ System. Rezultat testa prikazan u slučaju neuspješnog obavljanja postupka.

Tablica 5. Tumačenje rezultata

+: Došlo je do amplifikacije

-: Nije došlo do amplifikacije

1 Uzorak se smatra pozitivnim ako je dobivena Ct vrijednost manja od 40. Endogena interna kontrola (IC) može ili ne mora pokazivati signal amplifikacije. Ponekad detekcije interne kontrole nije potrebna jer veliki broj kopija cilja može uzrokovati preferencijalnu amplifikaciju ciljno-specifičnih nukleinskih kiselina.

2 Ako se samo jedno ciljno mjesto N gena amplificira, potvrđite sigmoidni oblik krivulje te intenzitet fluorescencije. U slučaju sumnjivog tumačenja, ovisno o dostupnom materijalu, također se preporučuje:

- a) ponovno ekstrahiranje i testiranje drugog alikvota istog uzorka (ako je moguće povećajte volumen uzorka na 750 µl) ili
- b) uzimanje novog uzorka te ponovno testiranje.

3 U slučaju negativnih ciljnih mjesta SARS-CoV-2, unutarnja kontrola mora pokazati signal amplifikacije s Ct manjim od 35. Ct vrijednosti mogla bi varirati zbog endogene interne kontrole te se radi o domaćinskom genu koji bi morao biti prisutan u svim ljudskom stanicama s jezgrom u izvornom uzorku. Ako postoji odsutno signalata ili Ct vrijednost ≥ 35 endogene interne kontrole, rezultat se smatra "Neriješenim" te je potrebno ponovno testiranje.



U slučaju stalnog dvosmislenog rezultata preporučuje se pregledati upute za uporabu te proces ekstrakcije kojeg koristi korisnik; za potvdu ispravne učinkovitosti svakog RT-qPCR koraka te pregled parametara; te za provjeru sigmoidnog oblika krivulje i intenzitet fluorescencije.

Rezultate testa treba procijeniti zdravstveni djelatnik uzimajući u obzir anamnezu, kliničke simptome i druge dijagnostičke testove.

10. Ograničenja testa

- Rezultate testa treba procijeniti zdravstveni djelatnik uzimajući u obzir anamnezu, kliničke simptome i druge dijagnostičke testove.
- Iako se ovaj test može koristiti s drugim vrstama uzorka, validiran je s nazofaringealnim/orofaringealnim brisovima prikupljenima u VTM.
- Za dobru učinkovitost testa liofilizirani proizvod mora biti na dnu epruvete te ne smije prianjati na gornjem dijelu epruvete ili čepa od folije. Lagano pritisnite svaku epruvetu na tvrdoj površini kako biste se uvjericili da je sav proizvod na dnu epruvete.
- Izgled reakcije smjese u stabiliziranom formatu koji se obično nalazi na dnu epruvete razlikuje se od uobičajenog (bez stožastog oblika, inhomogeni, manji/veći i/ili bojom različit od bjeličastog) ne mijenja funkcionalnost testa.
- Kvaliteta testa ovisi o kvaliteti uzorka; mora se ekstrahirati odgovarajuća nukleinska kiselina iz respiratornih uzorka.
- Ovaj test je samo kvalitativne test te ne osigurava kvantitativni vrijednosti te ne ukazuje na broj prisutnih organizama.
- Izuzetno niske razine ciljnog elementa ispod razine detekcije mogu biti detektirane, no moguće je da se rezultati neće moći ponoviti.
- Postoji mogućnost pojave lažnih pozitivnih rezultata zbog unakrsne kontaminacije izazvane uzorcima suspektnima na SARS-CoV-2 koji sadrže visoke koncentracije ciljne RNK ili kontaminacije izazvane proizvodima koji sadrže PCR iz ranijih reakcija.
- Kombinacije specifične početnice i sonde za detekciju uščuvanih regija N gena koji se koristi u VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System osmišljeni su na temelju američkog Centra za kontrolu bolesti za specifičnu detekciju SARS-CoV-2 amplifikacijom dvije jedinstvene regije N gena. Ne pokazuju značajnu kombiniranu homologiju s ljudskim genomom, ljudskom mikroflorom, SARS-CoV ili ostalim koronavirusima što bi moglo rezultirati u predvidivim lažno pozitivnim rezultatima.
- Lažno negativni rezultati mogu nastati uslijed nekoliko čimbenika te njihovih kombinacija uključujući:
 - nepravilno prikupljanje uzorka, transport, pohrana i/ili metode rukovanja.
 - Nepravilne postupke obrade (uključujući ekstrakciju RNK).
 - Degradacija virusne RNK tijekom otpreme/pohrane i/ili obrade uzorka.
 - Mutacije ili polimorfizmi na veznim regijama početnice ili sonde mogu utjecati na detekciju novih ili nepoznatih varijanti SARS-CoV-2.
 - Virusno opterećenje u uzorku koje je ispod granice detekcije za test.



- Prisutnost RT-qPCR inhibitora ili drugih tipova interferirajućih tvari. Nisu procijenjeni utjecaji cjepiva, antivirusnih terapija, antibiotika, kemoterapijskih sredstva ili imunosupresiva korištenih za prevenciju COVID-19 ili tijekom liječenja infekcije.
- Neuspjeh u pridržavanju uputa za uporabu te prilikom postupka testiranja.
- Amplifikacija pojedinačnog ciljnog mesta ili čak slučajno pozitivnih rezultata ukazuje na blago različito iskorištenje amplifikacije ciljnog mesta N gena. Uzorci s niskim virusnim opterećenjem mogu rezultirati u N pojedinačnoj ciljnoj amplifikaciji. U slučaju sumnji, preporučuje se обратити se referentnom laboratoriju za daljnje testiranje.
- Neki uzorci možda neće iskazati amplifikacijske krivulje RNase P zbog niskog broja ljudskih stanica u izvornom kliničkom uzorku. Negativni signal unutarnje kontrole ne isključuje prisutnost RNK virusa SARS-CoV-2 u kliničkom uzorku.
- Pozitivan rezultat testa ne ukazuje nužno na prisutnost vidljivih virusa te ne ukazuje da su ti virusi infektivni ili uzročni agensi za kliničke simptome. Međutim, pozitivni rezultat ukazuje na prisutnost ciljnih virusnih sekvenči (N gena).
- Negativni rezultati ne isključuju infekciju SARS-CoV-2 i ne smiju se koristiti kao jedini temelj za odlučivanje o liječenju ili pružanju druge zdravstvene skrbi bolesniku. Nisu određene optimalne vrste uzoraka i izračun vremena vršnih razina virusa tijekom infekcija čiji je uzrok SARS-CoV-2. Za detekciju virusa može biti potrebno prikupljanje više uzoraka (vrste i vremenske točke) od istog bolesnika.
- Ako su dijagnostički testovi za ostale respiratorne bolesti negativni, a klinička slika bolesnika te epidemiološke informacije sugeriraju da je moguća infekcija SARS-CoV-2, potrebno je razmotriti mogućnost lažnog negativnog rezultata i ponovnog testiranja bolesnika.
- U slučaju neriješenih, neutvrdnih ili nepotpunih rezultata primjenom kompleta za detekciju VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System potrebno je ponovno testiranje. Neriješeni rezultati mogu biti posljedica prisutnosti inhibitora u uzorku ili nepravilne rehidracije epruvete s liofiliziranom reakcijskom smjesom. Ako postoji kvar na instrumentu, dobivaju se neutvrdni ili nepotpuni rezultati.

11. Kontrola kvalitete

Komplet VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System sadrži endogenu unutarnju kontrolu (IC) u svakoj reakcijskoj epruveti pomoću koje se potvrđuje pravilno provođenje tehničke.

12. Radne karakteristike

12.1. Klinička osjetljivost i specifičnost

Klinička učinkovitost kompleta VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System testirana je uporabom 254 uzorka iz dišnog sustava (nazofaringelani brijevi u transportnom mediju Vircell) od pacijenata s kliničkom sumnjom na bolest COVID-19 ili drugu sličnu respiratornu bolest. Retrospektivno-komparativna analiza provedena je s kompletom VIASURE SARS-CoV-2 (N1 + N2) Real time PCR Detection Kit for BD MAX™ System te su ti rezultati uspoređeni s onima dobivenim s kliničkom dijagnozom dobivenom s testom Simplexa™ COVID-19 Direct assay s disrepantnom analizom provedenom s protokolom Charité.



	Alternativni testovi RT-PCR			
		+	-	Ukupno
+	63	2*	65	
-	0	189	189	
Ukupno	63	191	254	

Tablica 6. Komparativni rezultati za SARS-CoV-2.

*Početna dijagnoza za jedan ili dva uzorka nije bila valjana te je prijavljena pacijentu kao pozitivna u svrhu prevencije i razdoblja karantene.

VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System detektirala je dva pozitivna uzorka koji nisu detektirani uporabom testa Simplexa™ COVID-19 Direct assay te protokola Charité.

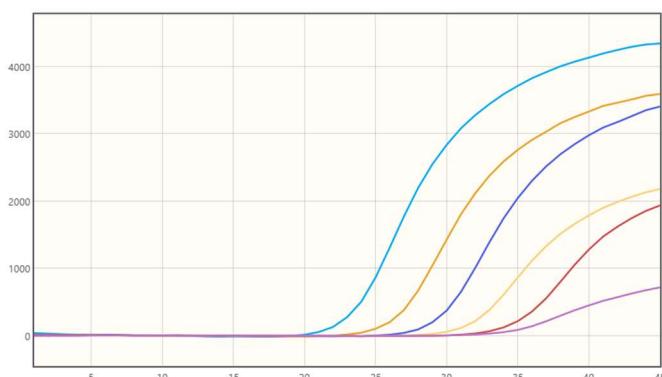
Postotak pozitivnog slaganja (PPA) te postpotak negativnog slaganja (NPA) za komplet za detekciju VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System su >99% odnosno 98%.

Rezultati pokazuju veliko slaganje za detekciju SARS-CoV-2 uporabom VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System.

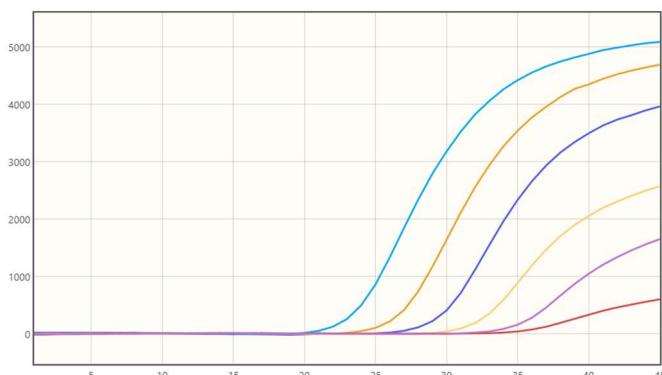
12.2. Analitička osjetljivost

Granica detekcije kompleta VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System iznosi ≥ 5 kopija genoma po reakciji s pozitivnom stopom od $\geq 95\%$.

Slika 2. Serije razrjeđivanja predloška SARS-CoV-2 (N1 + N2) (9.9×10^4 - 9.9×10^0 te 5.0×10^0 kopija genoma po reakciji) na sustavu BD MAX™ System (475/520 (FAM) kanal).



Slika 3. Serija razrjeđivanja predloška SARS-CoV-2 (N1 + N2) ($9,9 \cdot 10^4$ - $9,9 \cdot 10^0$ te $5,0 \cdot 10^0$ kopija genoma po reakciji) radi na sustavu BD MAX™ System (630/665 (Cy5) kanal).



12.3. Analitička specifičnost

Specifičnost testa SARS-CoV-2 (N1 + N2) potvrđena je testiranjem panela koji se sastoji od različitih mikroorganizama koji predstavljaju najčešće respiratorne patogene. Nije zabilježena unakrsna reaktivnost između bilo kojih od sljedećih testiranih mikroorganizama:

Testiranje unakrsne reaktivnosti					
Ljudski adenovirus tipovi 1-5, 8, 15, 31, 40 i 41	-	Influenza A/New Caledonia/20/99(H1N1) virus	-	Legionella longbeachae	-
Ljudski bokavirus	-	Influenza A/California/7/2009(H1N1)pdm09 virus	-	Legionella micdadei	-
Bordetella bronchiseptica	-	Influenza A/Michigan/45/2015 (H1N1)pdm09 virus	-	Legionella pneumophila	-
Bordetella holmesii	-	Influenza A/Singapore/GP1908/2015, IVR-180 (H1N1)pdm09 virus	-	Ljudski metapneumovirus A i B	-
Bordetella parapertussis	-	Influenza A/Victoria/210/2009 (H3N2) virus	-	Moraxella catarrhalis	-
Bordetella pertussis	-	Influenza A/Thüringen/5/2017 (H3N2) virus	-	Mycoplasma pneumoniae	-
Chlamydia caviae	-	Influenza A/Switzerland/9715293/2013 (H3N2) virus	-	Mycobacterium tuberculosis koja nije otporna na rifampicin	-
Chlamydia psittaci genotip A i C	-	Influenza A/Hong Kong/4801/2014, NYMC X-263B (H3N2) virus	-	Virusi ljudske parainfluence tipa 1, 2, 3 i 4	-
Chlamydophila pneumoniae CM-1	-	Influenza A/ South Australia/55/2014, IVR-175 (H3N2) virus	-	Pneumocytis jirovecii tip A1 i g885652	-
Ljudski koronavirus 229E, OC43, NL63 i HKU1	-	Influenza A/DE-SH/Reiherente/AR8444/ 2016 (H5N8) virus	-	Ljudski rinovirus tip C	-
MERS koronavirus	-	Influenza A/Anhui/1/2013 (H7N9) virus	-	Staphylococcus aureus podsoj aureus	-
SARS koronavirus soj Frankfurt 1	-	Influenza B/Brisbane/60/2008 virus	-	Staphylococcus epidermidis	-
Enterovirus 68 i 71	-	Influenza B/Florida/04/06 virus	-	Streptococcus pneumoniae Z2022	-
Enterovirus Echovirus 11 i 30	-	Influenza B/Phuket/3073/2013 virus	-	Streptococcus pyogenes	-
Enterovirus Coxsackievirus A24, A9 i B3	-	Legionella bozemanii	-	Streptococcus salivarius	-
Haemophilus influenzae Minna	-	Legionella dumoffii	-	Respiratori sincicijski virus (RSV) A i B	-

Tablica 7. Referentni patogeni mikroorganizmi korišteni u ovom ispitivanju.



12.4. Analitička reaktivnost

Reaktivnost VIASURE SARS-CoV-2 (N1 + N2) Real Time PCR Detection Kit for BD MAX™ System za SARS-CoV-2 procijenjena je naspram RNK iz ljudskog soja 2019-nCoV BetaCoV/Germany/BavPat1/2020 p.1, ljudski soj 2019-nCoV 2019-nCoV/Italy-INMI1, SARS-CoV-2 strain 2019nCoV/USA-WA1/2020, sintetska RNK kontrolira dvije varijante virusa SARS-CoV-2: MT007544.1 (SARS-CoV2 izolat Australia/VIC01/2020) i MN908947.3 (SARS-CoV-2 izolat Wuhan-Hu-1), i pokazala je pozitivne rezultate.

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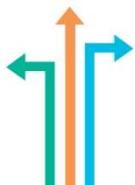
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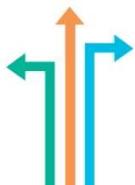
14. Symbols for IVD components and reagents/ Simboli za IVD komponente i reagense

IVD	In vitro diagnostic device In vitro dijagnostički uredaj		Keep dry Čuvati na suhom		Use by Rok valjanosti		Manufacturer Proizvođač	LOT	Batch code (Lot) Šifra serije
	Consult Instructions for Use Pogledajte upute za upotrebu		Temperature limitation Ograničenje temperature		Contains sufficient for <n> test Sadržaj dovoljan za <n> test(ova)	DIL	Sample diluent Razrjeđivač uzorka	REF	Catalognumber Kataloški broj

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