VIASURE

Real Time PCR Detection Kits

by CerTest BIOTEC

SARS-CoV-2 S gene

Handbook for the following references/

Priručnik za sljedeće reference/

VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit **BD REF 444212**

to be used with the BD MAX™ System

koristi se sa BD MAX™ sustavom

ENGLISH

1. Intended use

VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit is designed for the specific identification and differentiation of 2019 Novel Coronavirus (SARS-CoV-2) in respiratory samples from patients with signs and symptoms of COVID-19 infection. This test is intended to be used as an aid in the identification in the diagnosis of COVID-19 in combination with patient's clinical signs and symptoms and epidemiological risk factors. The assay uses the BD MAX™ System for automated extraction of RNA and subsequent real-time PCR employing the reagents provided combined with universal reagents and disposables for the BD MAX™ System. RNA from respiratory specimens is detected using fluorescent reporter dye probes specific for SARS-CoV-2.

2. Summary and Explanation

Coronavirus are enveloped non-segmented positive-sense RNA viruses and belong to *Coronaviridae* family [1,2]. There are six coronavirus species known to cause human diseases [2]. Four viruses (229E, OC43, NL63 and HKU1) cause common cold symptoms and the other two (severe acute respiratory syndrome coronavirus (SARS-CoV) and Middle East respiratory syndrome coronavirus (MERS-CoV)) are zoonotic and producing more severe complications [2]. SARS-CoV and MERS-CoV have caused more than 10,000 cumulative cases in the past two decades, with mortality rates of 34% MERS-CoV and 10% SARS-CoV [1,3].

In December 2019, some people that worked at or lived around the Huanan seafood market in Wuhan, Hubei Province, China, have presented pneumonia of unknown cause [2,4]. Deep sequencing analysis of the respiratory samples indicated a novel coronavirus, which was named firstly 2019 novel coronavirus (2019-nCoV) and lately SARS-CoV-2 [5].

Human-to-human transmission of the SARS-CoV-2 has been confirmed, even in the incubation period without symptoms, and the virus causes severe respiratory illness like those SARS-CoV produced [1,6,7]. Although the pneumonia is the principal illness associated, a few patients have developed severe pneumonia, pulmonary edema, acute respiratory distress syndrome, or multiple organ failure and death [1,4]. Centers of Disease Control and Prevention (CDC) believes that symptoms of SARS-CoV-2 may appear in as few as 2 days or as long as 14 days after exposure, being the most common fever, cough, myalgia and dyspnea [1,4,8]. Less common symptoms are sore throat, headache, diarrhea and vomiting [1,4]. It seems that older males with comorbidities have been more affected [4].

Diagnosis of SARS-CoV-2 is performed detecting conventional causes of pneumonia early and detected by next-generation sequencing or real-time RT-PCR methods [1,9]. Several assays that detect the SARS-CoV-2 have been are currently available, and listed on the WHO (World Health Organization) website https://www.who.int/emergencies/diseases/novel-coronavirus-2019/technical-guidance/laboratory-guidance [10].

WHO recommends upper respiratory tract specimens (nasopharyngeal and oropharyngeal swabs) and/or lower respiratory specimens (sputum, endotracheal aspirate, or bronchoalveolar lavage) for the identification of SARS-

CoV-2 [9,11]. In addition, other clinical specimens as blood, urine and stool may be collected to monitor the presence of the virus [9,11].

3. Principle of the procedure

VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit is designed for the identification of SARS-CoV-2 in respiratory samples. The detection is done in one step real time RT-PCR format where the reverse transcription and the subsequent amplification of specific target sequence occur in the same reaction well. The isolated RNA target is transcribed generating complementary DNA by reverse transcriptase which is followed by the amplification of a conserved region of *S* gene using specific primers and a fluorescent-labeled probe.

VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit is based on 5´ exonuclease activity of DNA polymerase. During DNA amplification, this enzyme cleaves the probe bound to the complementary DNA sequence, separating the quencher dye from the reporter. This reaction generates an increase in the fluorescent signal which is proportional to the quantity of the target template. This fluorescence is measured on the BD MAX™ System.

VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit contains in each tube all the components necessary for real time PCR assay (specific primers/probes, dNTPS, buffer, polymerase) in a stabilized format, as well as an internal control to monitor PCR inhibition. SARS-CoV-2 is amplified and detected in channel 475/520 and the internal control (IC) in in channel 530/565.

4. Reagents provided

VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection kit includes the following materials and reagents detailed in Table 1:

Reference	Reagent/Material	Description	Color	Amount
VS-NCO112	*SARS-CoV-2 S gene* reaction tube	A mix of enzymes, primers probes, buffer, dNTPs, stabilizers and internal control in stabilized format	Transparent Green foil	2 pouches of 12 tubes
VS-RB09	Rehydration Buffer tube	Solution to reconstitute the stabilized product	Transparent Orange foil	1 pouch of 24 tubes

Table 1. Reagents and materials provided in VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection kit with Ref. VS-NCO124 (444212).

5. Reagents and equipment to be supplied by the user

The following list includes the materials and equipment that are required for use but not included in the VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection kit.

- Real-time PCR instrument: BD MAX™ System.
- BD MAX™ ExK™ TNA-3 (Ref: 442827 or 442828).
- BD MAX™ PCR Cartridges (Ref: 437519).
- Vortex.
- Micropipettes (accurate between 2 and 1000 µL).

- Filter tips.
- Powder-free disposable gloves.

6. Transport and storage conditions

- The kits can be shipped and stored at 2-40°C until the expiration date which is stated on the label.
- After opening the aluminum pouches which contain the reaction tubes, they can be used for up to 28 days.

7. Precautions for users

- The product is indented for use by professional users only, such as laboratory or health professionals and technicians, trained in molecular biological techniques.
- For *in vitro* diagnostic use.
- Do not use expired reagents and/or materials.
- Do not use the kit if the label that seals the outer box is broken.
- Do not use reagents if the protective box is open or broken upon arrival.
- Do not use reagents if the protective pouches are open or broken upon arrival.
- Do not use reagents if desiccant is not present or broken inside reagent pouches.
- Do not remove desiccant from reagent pouches.
- Close protective pouches of reagents promptly with the zip seal after each use. Remove any excess air in the pouches prior to sealing.
- Do not use reagents if the foil has been broken or damaged.
- Do not mix reagents from different pouches and/or kits and/or lots.
- Protect reagents from humidity. Prolonged exposure to humidity may affect product performance.
- Keep components away from light.
- In cases where other PCR tests are conducted in the same general area of the laboratory, care must be taken to ensure that the VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection kit, BD MAX™ ExK™ TNA-3 extraction kit, any additional reagents required for testing, and the BD MAX™ System are not contaminated. Avoid microbial and ribonuclease (RNase)/deoxyribonuclease (DNase) contamination of reagents at all times. The use of sterile RNase/DNase-free disposable aerosol resistant or positive displacement pipette tips is recommended. Use a new tip for each specimen. Gloves must be changed before manipulating reagents and cartridges.
- To avoid contamination of the environment by amplicons, do not break apart the BD MAX™ PCR Cartridge after use. The seals of the BD MAX™ PCR Cartridge are designed to prevent contamination.
- Design a unidirectional workflow. It should begin in the Extraction Area and then move to the Amplification and Detection Area. Do not return samples, equipment and reagents to the area in which the previous step was performed.
- Follow Good Laboratory Practices. Wear protective clothing, use disposable gloves, goggles and mask. Do not eat, drink or smoke in the working area. Wash your hands after finishing the test.
- Samples must be treated as potentially infectious as well as all the reagents and materials that have been exposed to the samples and they must be handled according to the national safety regulations. Take necessary precautions during the collection, storage, treatment and disposal of samples.

- Regular decontamination of commonly used equipment is recommended, especially micropipettes and work surfaces.
- Consult the BD MAX™ System User's Manual for additional warnings, precautions and procedures.

8. Test procedure

8.1. SAMPLE COLLECTION, STORAGE AND TRANSPORT

The VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection kit has been validated on negative nasopharyngeal/ oropharyngeal swab collected in viral transport media (VTM) Vircell S.L., Spain) and nucleic acid isolated from positive nasopharyngeal/ oropharyngeal swab collected in VTM.

Another different types of samples from nasopharyngeal/oropharyngeal swabs in VTM must be validated by the user.

Collection, storage and transport specimens should be maintained per the conditions validated by the user. Overall, respiratory samples should be collected and labelled appropriately in clean containers with or without transport media (depending on sample type), and processed as soon as possible to guarantee the quality of the test. The specimens should be transported at 2 to 8°C for up to 48 hours, following the local and national regulations for the transport of pathogen material. For long term transport (more than 48 hours), we recommend shipping at ≤-20°C. It is recommended to use fresh specimens for the test. The samples can be stored at 2 to 8°C for up to 48 hours or frozen at -20°C or ideally at -70°C for conservation. Repeated freeze-thaw cycles should be avoided in order to prevent degradation of the sample and nucleic acids.

8.2. SAMPLE PREPARATION AND RNA EXTRACTION

Perform the sample preparation according to the recommendations in the instructions for use of the extraction kit used, BD MAX™ ExK™ TNA-3. Note that some other samples may require pre-processing. Application-specific extraction preparation procedures should be developed and validated by the user.

1. Pipette between 200 and 750 µL of nasopharyngeal/ oropharyngeal swab collected in viral transport media (VTM) into a BD MAX™ TNA-3 Sample Buffer Tube and close the tube with a septum cap. Ensure complete mixing by vortexing the sample at high speed for 1 minute. Proceed to BD MAX™ System Operation.

8.3. PCR PROTOCOL

Note: Please, refer to the BD MAX™ System User's Manual for detailed instructions.

8.3.1. Creating PCR test program for VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection kit

Note: If you have already created the VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection test, you can skip step 8.3.1 and go directly to 8.3.2.

1) On the "Run" screen of the BD MAX™ System, select the "Test Editor" tab.

2) Click the "Create" button.

3) In the "Test Name" window, name your test: i.e. VIASURE *SARS-CoV-2 S gene*.

4) In the "Extraction Type" drop down menu, select "ExK TNA-3".

5) In the "Master Mix Format" drop down menu, choose "Type 5".

   a. Note: Product may be used in combination with an additional VIASURE for BD MAX test, then select "Dual Master Mix Concentrated Lyophilized MM with Rehydration Buffer (Type 5)".

6) In the "Sample extraction parameters" select "User defined" and adjust sample volume to the volume of clinical specimen used plus 500 µL.

   a. Example: If pipette 200 µL of respiratory clinical specimen into a BD MAX TNA-3 Sample Buffer Tube then set parameter to 700 µL.

   b. Note: maximum setting is 950 µL

7) In the "Ct Calculation" select "Call Ct at Threshold Crossing".

8) In "PCR settings" tab enter the following parameters: "Channel Settings", "Gains" and "Threshold" (Table 2).

   a. Note: Product may be used in combination with an additional VIASURE for BD MAX test, PCR Settings and Test Steps should be completed for snap 2 (green) and snap 4 (blue) positions.

Channel	Alias	Gain	Threshold	Ct Min	Ct Max
475/520 (FAM)	SARS-CoV-2 *S* gene	60	100	0	40
530/565 (HEX)	IC	80	100	0	40
585/630 (ROX)	-	0	0	0	0
630/665 (Cy5)	-	0	0	0	0
680/715 (Cy5.5)	-	0	0	0	0

Table 2. PCR settings.

Note: It is recommended to set the minimum threshold values listed above for each channel as a starting point, but the final settings must be determined by the end-user during the result interpretation in order to ensure that thresholds fall within the exponential phase of the fluorescence curves and above any background signal. The threshold value for different instruments may vary due to different signal intensities.

9) In "PCR settings" tab enter the following parameters "Spectral Cross Talk" (Table 3), as well

		False Receiving Channel				
	Channel	475/520	530/565	585/630	630/665	680/715
Excitation Channel	475/520	-	0.0	0.0	0.0	0.0
	530/565	0.0	-	0.0	0.0	0.0
	585/630	0.0	0.0	-	0.0	0.0
	630/665	0.0	0.0	0.0	-	0.0
	680/715	0.0	0.0	0.0	0.0	-

Table 3. Spectral cross-talk parameters.

10) In "Test Steps" tab, enter the PCR protocol (Table 4).

Step Name	Profile Type	Cycles	Time (s)	Temperature	Detect
Reverse transcription	Hold	1	900	45°C	-
Initial denaturation	Hold	1	120	98°C	-
Denaturation and Annealing/Extension (Data collection)	2-Temperature	45	10	95°C	-
			58	60°C	✓

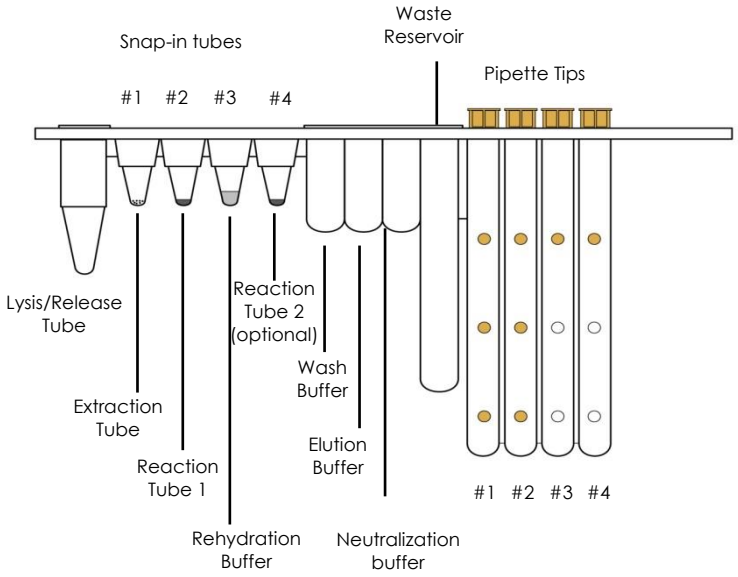
Table 4. PCR protocol.

11) Click the "Save Test" button.

8.3.2. BD MAX™ Rack set up

1) For each sample to be tested, remove one Unitized Reagent Strips from the BD MAX™ ExK TNA-3 kit. Gently tap each strip onto a hard surface to ensure that all the liquids are at the bottom of the tubes and load on the BD MAX™ System sample racks.
2) Remove the required number of BD MAX™ ExK TNA Extraction Tubes (B4) (white foil) from their protective pouch. Snap the Extraction Tube(s) (white foil) into its corresponding positions in the TNA strip (Snap position 1, white color coding on the rack. See Figure 1). Remove excess air, and close the pouch with the zip seal.
3) Determine and separate the appropriate number of VIASURE *SARS-CoV-2 S gene* reaction tube (green foil) and snap into their corresponding positions in the strip (Snap position 2, green color coding on the rack. See Figure 1).
    a. Remove excess air, and close aluminum pouches with the zip seal.
    b. In order to carry out a correct rehydration, please make sure that the lyophilized product is in the bottom of the tube and is not adhered to the top area of the tube or to the foil seal.
        i. Note: If you choose the format "Dual Master Mix Concentrated Lyophilized MM with Rehydration Buffer (Type 5)" (Section 8.3.1), determine and separate the appropriate number of additional VIASURE reaction tubes (different foil) and snap into their corresponding positions in the strip (Snap position 4, blue color coding on the rack. See Figure 1). Remove excess air, and close aluminum pouches with the zip seal.
4) Remove the required number of Rehydration Buffer tubes (orange foil) and snap into their corresponding positions in the strip (Snap position 3, non-color coding on the rack. See Figure 1). Remove excess air, and close the pouch with the zip seal.
    a. In order to ensure a correct transfer, please make sure that the liquid is in the bottom of the tube and is not adhered to the top area of the tube or to the foil seal.

Figure 1. BD MAX™ TNA Reagent Strip (TNA) from the BD MAX™ ExK TNA-3 kit.



8.3.3. BD MAX™ Instrument set up

1) Select the "Work List" tab on the "Run" screen of the BD MAX™ System software v4.50A or higher.
2) In the "Test" drop down menu, select VIASURE *SARS-CoV-2 S gene* (if not already created see Section 8.3.1).
3) Select the appropriate kit lot number (found on the outer box of extraction kit used) from the pull down menu (optional).
4) Enter the Sample Buffer Tube identification number into the Sample tube window of the Worklist, either by scanning the barcode with the scanner or by manual entry.
5) Fill the Specimen/Patient ID and/or Accession window of the Worklist and click the "Save" button. Continue until all Sample Buffer Tubes are entered. Ensure that the specimen/patient ID and the Sample Buffer Tubes are accurately matched.
6) Place the prepared Sample Buffer Tube into the BD MAX™ Rack(s).
7) Load the rack(s) into the BD MAX™ System (Rack A is positioned on the left side of the BD MAX™ System and Rack B on the right side).
8) Place the required number of BD MAX™ PCR Cartridge(s) into the BD MAX™ System.
9) Close the BD MAX™ System door.
10) Click "Start Run" to begin the procedure.

8.3.4 BD MAX™ report

1) In main menu, click the "Results" button.
2) Either double click on your run in the list or press the "view button".
3) Click on "Print", select: "Run Details, Test Details and Plot…"
4) Click on "Print or Export button" on the "Run Reports" screen.

9. Result interpretation

For a detailed description on how to analyze data, refer to the BD MAX™ System User's manual.

The analysis of the data is done by the BD MAX™ software according to the manufacturer´s instructions. The BD MAX™ software reports Ct values and amplification curves for each detector channel of each sample tested in the following way:

- Ct value of 0 indicates that there was no Ct value calculated by the software with the specified Threshold (see Table 2). Amplification curve of the sample showing a "0" Ct value must be checked manually.

- Ct value of -1 indicates that no amplification process has occurred.

- Any other Ct value should be interpreted in correlation with the amplification curve and according to the sample interpretation guidelines outlined in Table 5.

Check Internal Control signal to verify the correct functioning of the amplification mix. In addition, check that there is no report of BD MAX™ System failure.

-Results should be read and analyzed using the following table:

SARS-CoV-2 *S* gene (475/520)	**Internal control (530/565)**	**Interpretation**
-	+	**SARS-CoV-2 *S* gene RNA Not Detected**
+	+/-	**SARS-CoV-2 *S* gene RNA Detected**
-	-	**Unresolved (UNR) Result obtained in the presence of inhibitors in the PCR reaction or when a general problem (not reported by an error code) with the sample processing and/or amplification steps occurs.**
IND	IND	**Indeterminate assay result (IND). Due to BD MAX™ System failure. Assay result displayed in case of an instrument failure linked to an error code.**
INC	INC	**Incomplete assay result (INC). Due to BD MAX™ System failure. Assay result displayed in case of failure to complete run.**

Table 5. Sample interpretation

+: Amplification occurred

-: No amplification occurred

A sample is considered positive if the Ct value obtained is less than 40. The internal control may or may not show an amplification signal because a high copy number of target can cause preferential amplification of target-specific nucleic acids instead of the internal control. In these cases, the detection of the IC is not necessary.

A sample is considered negative, if the sample shows no amplification signal in the detection system but the internal control is positive. An inhibition of the PCR reaction can be excluded by the amplification of internal control.

In case of unresolved results (UNR), absence of internal control signal in negative sample it is recommended to repeat the assay diluting the sample 1:10 to check for possible problems of inhibition.

The results of the test should be evaluated by a health care professional in the context of medical history, clinical symptoms and other diagnostic tests.

10. Limitations of the test

- The results of the test should be evaluated by a health care professional in the context of medical history, clinical symptoms and other diagnostic tests.
- Although this assay can be used with other types of samples it has been validated with nasopharyngeal/ oropharyngeal swab collected in VTM.
- The quality of the test depends on the quality of the sample; proper extracted nucleic acid from respiratory samples must be extracted.
- Extremely low levels of target below the limit of detection might be detected, but results may not be reproducible.
- There is a possibility of false positive results due to cross-contamination by SARS-CoV-2 suspicious samples containing high concentrations of target RNA or contamination due to PCR products from previous reactions.
- The specific primer and probe combination for detection of the *S* gene used in VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit designed for the detection of SARS-CoV-2, do not show significant combined homologies with the human genome, human microflora, SARS-CoV or other coronaviruses, which might result in predictable false positive.
- False Negative results may arise from several factors and their combinations, including:
    - Improper specimens' collection, transport, storage, and/or handling methods.
    - Improper processing procedures (including RNA extraction).
    - Degradation of the viral RNA during sample shipping/storage and/or processing.
    - Mutations or polymorphisms in primer or probe binding regions may affect detection of new or unknown SARS-CoV-2 variants.
    - A viral load in the specimen below the limit of detection for the assay.
    - The presence of RT-qPCR inhibitors or other types of interfering substances. The impacts of vaccines, antiviral therapeutics, antibiotics, chemotherapeutics or immunosuppressant drugs used to prevent COVID-19 or used during the treatment of the infection have not been evaluated.
    - Failure to follow instructions for use and the assay procedure.
- Negative results do not preclude SARS-CoV-2 infection and should not be used as the sole basis for treatment or other patient management decisions. Optimum specimen types and timing for peak viral levels during infections caused by SARS-CoV-2 have not been determined. The collection of multiple specimens (types and time points) from the same patient may be necessary to detect the virus.
- If diagnostic tests for other respiratory illnesses are negative and the patient's clinical presentation and epidemiological information suggest that SARS-CoV-2 infection is possible, then a false negative result should be considered, and a re-testing of the patient should be discussed.

- A negative result does not preclude the presence of SARS-CoV-2 virus RNA in a clinical specimen. If clinical observations, patient history and epidemiological information suggest COVID-19 infection, re-testing increasing sample volume should be considered.
- In the case of obtaining Unresolved, Indeterminate or Incomplete results using VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit retesting will be required. Unresolved results may be due to the presence of inhibitors in the sample or an incorrect rehydration of lyophilized reaction mix tube. If there is an instrument failure, Indeterminate or Incomplete results will be obtained.

11. Quality control

VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit contains an internal control (IC) in each reaction tube which confirms the correct performance of the technique.

12. Performance characteristics

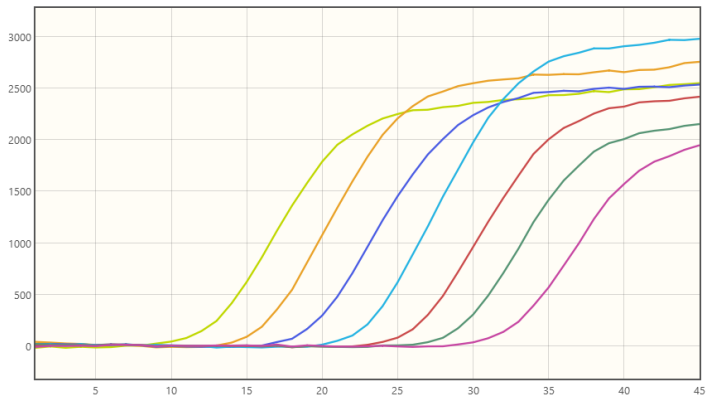
12.1. Clinical sensitivity and specificity

The clinical performance of VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit was tested using 4 nucleic acids isolated from positive nasopharyngeal and/or oropharyngeal swabs collected in VTM and 15 respiratory samples (nasopharyngeal and/or oropharyngeal swabs in VTM) from patients with clinical suspicion of COVID-19 disease or other similar respiratory diseases. Four 4 SARS-CoV-2 positive samples were found and these results were in agreement with a PCR test developed according to the China CDC Primers and probes for detection 2019-nCoV. Additionally, the 4 SARS-CoV-2 positive samples were confirmed using a molecular detection method by the Spanish National Reference Center (Institute of Health Carlos III (ISCIII))(the protocol "2019-nCoV by real-time RT-PCR" suggested by Charité (Berlin), with modifications).
In conclusion, the results show a high sensitivity and specificity to detect SARS-CoV-2 using VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit.

12.2. Analytical sensitivity

VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit has a detection limit of ≥ 24 cDNA copies per reaction (cp/rxn) with a positive rate of ≥ 95%.

Figure 2. Dilution series of SARS-CoV-2 *S* gene ($2.4*10^7$-$2.4*10^1$ cp/rxn) template run on the BD MAX™ System (475/520 (FAM) channel).



12.3. Analytical specificity

The specificity of the SARS-CoV-2 *S* gene assay was confirmed by testing a panel consisting of different microorganisms representing the most common respiratory pathogens. No cross-reactivity was detected between any of the following microorganisms tested:

Cross-reactivity testing					
Human Adenovirus types 1-5, 8, 15, 31, 40 and 41	-	Influenza A/California/7/2009(H1N1)pdm09-like virus	-	*Legionella dumoffii*	-
Human Bocavirus	-	Influenza A/Michigan/45/2015 (H1N1)pdm09 virus	-	*Legionella longbeachae*	-
Bordetella bronchiseptica	-	Influenza A/Singapore/GP1908/2015, IVR-180 (H1N1)pdm09 virus	-	*Legionella micdadei*	-
Bordetella holmesii	-	Influenza A/Victoria/210/2009 (H3N2) virus	-	*Legionella pneumophila*	-
Bordetella parapertussis	-	Influenza A/Thüringen/5/2017 (H3N2) virus	-	Human metapneumovirus A and B	-
Bordetella pertussis	-	Influenza A/Switzerland/9715293/2013 (H3N2) virus	-	*Moraxella catarrhalis*	-
Chlamydia caviae	-	Influenza A/Hong Kong/4801/2014, NYMC X-263B (H3N2) virus	-	*Mycoplasma pneumoniae*	-
Chlamydia psittaci genotype A and C	-	Influenza A/ South Australia/55/2014, IVR-175 (H3N2) virus	-	*Mycobacterium tuberculosis*	-
Chlamydophila pneumoniae	-	Influenza A/DE-SH/Reiherente/AR8444/ 2016 (H5N8) virus	-	Human parainfluenza 1, 2, 3 and 4 viruses	-
Human coronavirus 229E, OC43, NL63 and HKU1	-	Influenza A/Anhui/1/2013 (H7N9) virus	-	*Pneumocytis jirovecii* Type A1 and g885652	-
MERS Coronavirus	-	Influenza B/Brisbane/60/2008 virus	-	Human rhinovirus type C	-
SARS Coronavirus Strain Frankfurt 1	-	Influenza B/Florida/04/06 virus	-	*Staphylococcus aureus* subsp. *aureus*	-
Haemophilus influenzae MinnA	-	Influenza B/Phuket/3073/2013 virus	-	*Streptococcus pneumoniae*	-
Influenza A/New Caledonia/20/99(H1N1) virus	-	*Legionella bozemanii*	-	Respiratory syncytial virus (RSV) A and B	-

Table 6. Reference pathogenic microorganisms used in this study.

12.4. Analytical reactivity

The reactivity of VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit was evaluated against RNA from Human 2019-nCoV strain BetaCoV/Germany/BavPat1/2020 p.1, Human 2019-nCoV strain 2019-nCoV/Italy-INMI1, synthetic RNA controls for two variants of the SARS-CoV-2 virus: MT007544.1 (SARS-CoV2 isolate Australia/VIC01/2020) and MN908947.3 (SARS-CoV-2 isolate Wuhan-Hu-1), showing positive result.

HRVATSKI

1. Namjena

Komplet VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit dizajniran je za specifičnu identifikaciju i diferencijaciju novog koronavirusa 2019 (SARS-CoV-2) u respiratornim uzorcima bolesnika sa znakovima i simptomima infekcije COVID-19. Predviđeno je da se ovaj test koristi kao pomoć u dijagnosticiranju COVID-19 u kombinaciji s kliničkim znakovima i simptomima pacijenta te s epidemiološkim faktorima rizika. Test koristi BD MAX™ sustav za automatiziranu ekstrakciju RNK, a zatim lančanu reakciju polimeraze (PCR) u stvarnom vremenu s priloženim reagensima u kombinaciji s univerzalnim reagensima i jednokratnim materijalom za BD MAX™ sustav. Detekcija RNK iz respiratornih uzoraka vrši se primjenom sondi s fluorescentnim indikacijskim bojilom specifičnim za SARS-CoV-2.

2. Sažetak i objašnjenje

Koronavirusi su grupa ovijenih, nesegmentiranih, pozitivno usmjerenih RNK virusa koji spadaju u obitelj *Coronaviridae* [1,2]. Poznato je šest vrsta koronavirusa koji uzrokuju bolesti u ljudi [2]. Četiri virusa (229E, OC43, NL63 i HKU1) uzrokuju uobičajene simptome prehlade, a preostala dva (koronavirus teškog akutnog respiratornog sindroma (SARS-CoV) i koronavirus respiratornog sindroma Bliskog istoka (MERS-CoV)) su zoonotični i izazivaju teže komplikacije [2]. SARS-CoV i MERS-CoV prouzročili su preko 10.000 kumulativnih slučajeva u protekla dva desetljeća, sa stopama smrtnosti od 34% MERS-CoV i 10% SARS-CoV [1,3].

U prosincu 2019, nekoliko osoba koje su radile i živjele oko tržnice morskih plodova i životinja Huanan u Wuhanu, kineskoj pokrajini Hubei, oboljele su od upale pluća nepoznatog uzroka [2,4]. Analiza dubokog sekvenciranja respiratornih uzoraka ukazala je na novi koronavirus, koji je prvo dobio ime novi koronavirus 2019 (2019-nCoV), a zatim SARS-CoV-2 [5].

Potvrđen je prijenos s čovjeka na čovjeka, čak i tijekom inkubacijskog razdoblja bez simptoma, a virus uzrokuje teške respiratorne bolesti poput onih izazvanih virusom SARS-CoV [1,6,7]. Iako je upala pluća najčešća povezana bolest, u nekoliko bolesnika razvila se teška upala pluća, akutni respiratorni distres sindrom ili zakazivanje više organa i smrt [1,4]. Centri za kontrolu i prevenciju bolesti (CDC) iz SAD-a smatraju da se simptomi SARS-CoV-2 mogu pojaviti od 2 do 14 dana nakon izlaganja, pritom su najčešći vrućica, kašalj, mijalgija i zaduha[1,4,8]. Manje česti simptomi su grlobolja, glavobolja, proljev i povraćanje [1,4]. Čini se da su stariji muškarci s komorbiditetom pogođeniji [4].

Dijagnosticiranje SARS-CoV-2 provodi se ranom detekcijom konvencionalnih uzroka upale pluća i detektira se sekvenciranjem sljedeće generacije ili metodama lančane reakcije polimeraze reverznom transkripcijom (RT-PCR) u stvarnom vremenu [1,9]. Trenutno je dostupno nekoliko testova koji detektiraju SARS-CoV-2, a navedeni su na web-mjestu Svjetske zdravstvene organizacije (WHO) https://www.who.int/emergencies/diseases/novel-coronavirus-2019/technical-guidance/laboratory-guidance [10].

WHO preporučuje uzorke iz gornjih dišnih puteva (nazofaringealni i orofaringealni brisovi) i/ili donjih dišnih puteva (ispljuvak, endotrahealni aspirat, ili bronhoalveolarni lavaž) za identifikaciju SARS-CoV-2 [9,11]. Pored toga, mogu se prikupiti i drugi klinički uzorci poput krvi, mokraće i stolice za nadzor prisutnosti virusa [9,11].

3. Načelo postupka

Komplet VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit dizajniran je za identifikaciju SARS-CoV-2 u respiratornim uzorcima. Detekcija se obavlja u obliku lančane reakcije polimeraze reverznom transkripcijom (RT-PCR) u stvarnom vremenu koja se sastoji od jednog koraka, gdje se reverzna transkripcija i potom amplifikacija specifične ciljne sekvence vrši u istoj reakcijskoj jažici. Nakon izolacije RNK, vrši se njena transkripcija čime se dobiva komplementarna DNK zahvaljujući reverznoj transkriptazi, nakon koje slijedi amplifikacija konzervirane regije gena S primjenom specifičnih početnica i fluorescentno obojane sonde.

Komplet VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit temelji se na djelovanju 5' egzonukleaze u DNK polimerazi. Tijekom amplifikacije DNK ovaj enzim rascjepljuje sondu vezanu za sekvencu komplementarne DNK, čime se razdvaja supresorsku boju od indikacijskog gena. Ova reakcija dovodi do povećanja fluorescencijskog signala srazmjernog količini ciljnog predloška. Ta fluorescencija mjeri se na BD MAX™ sustavu.

Komplet VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit sadrži u svakoj epruveti sve potrebne komponente za obavljanje testa PCR-a (specifične početnice/sonde, dNTPS pufer, polimerazu) u stabiliziranom obliku, kao i unutarnju kontrolu za nadzor inhibicije PCR-a. SARS-CoV-2 je pojačan i otkriven u kanalu 475/520, a unutarnja kontrola (IC) u kanalu 530/565.

4. Reagensi koji se isporučuju

Komplet VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit sadrži sljedeće materijale i reagense navedene u tablici 1:

Referenca	Reagens/materijal	Opis	Boja	Količina
VS-NCO112	*SARS-CoV-2 S gene* reaction tube	Smjesa enzima, sondi/početnica, pufera, dNTP-ova, stabilizatora i unutarnje kontrole u stabiliziranom obliku	Prozirna zelena folija	2 vrećice s 12 epruveta
VS-RB09	Rehydration Buffer tube	Otopina za rekonstituciju stabiliziranog proizvoda	Prozirna narančasta folija	1 vrećica s 24 epruvete

Tablica 1. Reagensi i materijali koji se nalaze u kompletu VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit s referentnim brojem VS-NCO124 (444212).

5. Reagensi i oprema koju mora pribaviti korisnik

Sljedeći popis uključuje materijale i opremu koja je neophodna za korištenje, ali nije dio kompleta VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit.

- Instrument za lančanu reakciju polimeraze (PCR) u stvarnom vremenu: BD MAX™ System.
- BD MAX™ ExK™ TNA-3 (Ref: 442827 ili 442828).
- BD MAX™ PCR Cartridges (Ref: 437519).
- Vortex.
- Mikropipete (precizne u rasponu od 2 do 1000 µl).

- Nastavci za filtar.
- Nenaprašene jednokratne rukavice.

6. Uvjeti za transport i pohranu

- Kompleti se mogu transportirati i čuvati na temperaturi od 2-40°C do isteka roka valjanosti navedenog na naljepnici.
- Nakon otvaranja aluminijskih vrećica koje sadrže reakcijske epruvete mogu se iskoristiti u roku od 28 dana.

7. Mjere opreza za korisnike

- Proizvod je namijenjen isključivo uporabi od strane profesionalnih korisnika, kao što su laboratorijski ili zdravstveni djelatnici i tehničari, koji su prošli edukaciju o tehnikama molekularne biologije.
- Samo za *in vitro* dijagnostičku upotrebu.
- Nemojte koristiti reagense i/ili materijale kojima je istekao rok valjanosti.
- Nemojte koristiti komplet ako je potrgana naljepnica kojom je zatvorena vanjska kutija.
- Nemojte koristiti reagense ako je zaštitna kutija otvorena ili oštećena po prispijeću.
- Nemojte koristiti reagense ako su zaštitne vrećice otvorene ili oštećene po prispijeću.
- Nemojte koristiti reagense ako desikant u vrećicama s reagensima nije prisutan ili je oštećen.
- Nemojte vaditi desikant iz vrećica s reagensima.
- Nakon svake upotrebe odmah zatvorite zaštitne vrećice s reagensima pomoću patentnog zatvarača. Prije zatvaranja istisnite višak zraka iz vrećica.
- Nemojte koristiti reagense ako je folija potrgana ili oštećena.
- Nemojte miješati reagense iz različitih vrećica, kompleta i/ili serija.
- Zaštitite reagense od vlage. Duže izlaganje vlazi može utjecati na učinak proizvoda.
- Čuvajte komponente dalje od svjetlosti.
- U slučajevima gdje se drugi testovi PCR-om obavljaju u istom općem prostoru laboratorija potreban je oprez kako ne bi došlo do kontaminacije kompleta VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit, kompleta BD MAX™ ExK™ TNA-3 extraction kit, svih dodatnih reagensa potrebnih za testiranje i BD MAX™ sustava. Svakako izbjegavajte kontaminaciju reagensa s mikrobima i ribonukleazama (Rnase)/deoksiribonukleazama (DNase). Preporučuje se uporaba jednokratnih, sterilnih vrhova volumetrijskih pipeta ili otpornih na aerosol, bez Rnase/Dnase. Koristite novi vrh sa svakim uzorkom. Rukavice se moraju promijeniti prije rukovanja reagensima i patronama.
- Radi sprječavanja kontaminacije okruženja amplikonima, nemojte odvajati BD MAX™ PCR Cartridge nakon uporabe. Brtve na BD MAX™ PCR Cartridge dizajnirane su za sprječavanje kontaminacije.
- Koristite jednosmjeran radni proces. Treba započeti u području za ekstrakciju, a zatim preći u područje za amplifikaciju i detekciju. nemojte vraćati uzorke, opremu u reagense u područje u kojem je obavljen prethodni korak.
- Pridržavajte se dobre laboratorijske prakse. Nosite zaštitnu odjeću, jednokratne rukavice, zaštitne naočale i masku. Nemojte jesti, piti niti pušiti u radnom prostoru. Operite ruke nakon što završite test.

- Uzorci se moraju smatrati potencijalno zaraznim, a isto vrijedi za reagense i materijale koji su bili izloženi uzorcima i mora se rukovati u skladu s nacionalnim sigurnosnim propisima. Poduzmite neophodne mjere opreza prilikom prikupljanja, pohrane, tretiranja i odlaganja uzoraka u otpad.
- Preporučuje se redovita dekontaminacija opreme koja se često koristi, naročito mikropipeta i radnih površina.
- Posavjetujte se s Korisničkim priručnikom za BD MAX™ sustav u vezi s dodatnim upozorenjima, mjerama opreza i postupcima.

8. Testni postupak

8.1. PRIKUPLJANJE, POHRANA I TRANSPORT UZORAKA

Komplet VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit validiran je na negativnim nazofaringealnim/orofaringealnim brisovima prikupljenima u virusnom transportnom mediju (VTM) (Vircell S.L., Španjolska) i na nukleinskim kiselinama izoliranima iz pozitivnih nazofaringealnih/orofaringealnih brisova prikupljenima u VTM-u.

Drukčije vrste uzoraka iz nazofaringealnih/orofaringealnih brisova u VTM-u mora validirati korisnik.

Prikupljanje, pohrana i transport uzoraka treba obavljati u uvjetima koje je validirao korisnik. U globalu, respiratorne uzorke treba prikupiti i označiti pravilno u čiste spremnike sa ili bez transportnog medija (ovisno o vrsti uzorka) i obraditi što je moguće prije u cilju osiguranja kvalitete testa. Uzorci se trebaju transportirati na temperaturi od 2 °C do 8 °C tijekom 48 sati u skladu s lokalnim i nacionalnim propisima za transport patogenih materijala. Za dugotrajni transport (duži od 48 sata) preporučujemo otpremanje na temperaturi od ≤-20 °C. Preporučuje se upotreba svježih uzoraka za test. Uzorci se mogu čuvati na temperaturi od 2 °C do 8 °C tijekom 48 sati ili držati smrznuti na temperaturi od -20°C ili idealno na -70°C za pohranu. Treba izbjegavati ponavljanje ciklusa smrzavanja i odmrzavanja kako bi se spriječilo propadanje uzorka i nukleinskih kiselina.

8.2. PRIPREMA UZORAKA I EKSTRAKCIJA RNK

Prikupljanje uzorka obavite u skladu s preporukama iz uputa za upotrebu kompleta za ekstrakciju koji koristite, BD MAX™ ExK™ TNA-3. Moguće je da će neki drugi uzorci zahtijevati predobradu. Korisnik treba razviti i validirati postupke pripreme za ekstrakciju specifične za predmetnu primjenu.

1. Pipetom prenesite između 200 µl i 750 µl nazofaringealnog/orofaringealnog brisa prikupljenog u virusnom transportnom mediju (VTM) u BD MAX™ TNA-3 Sample Buffer Tube epruvetu za uzorak s puferom te je zatvorite čepom s membranom. Izmiješajte uzorak vrtloženjem pri visokoj brzini tijekom 1 minute. Pređite na rad sa BD MAX™ System Operation.

8.3. PROTOKOL ZA PCR

Napomena: Detaljne upute potražite u Korisničkom priručniku za MAX™ sustav.

8.3.1. Kreiranje programa za testiranje PCR-om za komplet VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit

Napomena: Ako ste već kreirali test za komplet VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit, možete preskočiti korak 8.3.1 i preći izravno na 8.3.2.

1) Na zaslonu „Run" (Pokreni) na BD MAX™ sustavu odaberite karticu „Test Editor" (Uređivač testa).
2) Kliknite na gumb „Create" (Kreiraj).
3) U prozoru „Test Name" (Naziv testa) dajte ime svom testu, tj. VIASURE *SARS-CoV-2 S gene*.
4) S padajućeg izbornika „Extraction Type" (Vrsta ekstrakcije) odaberite „ExK TNA-3".
5) S padajućeg izbornika „Master Mix Format" (Oblik glavne smjese) odaberite „Type 5" (Tip 5).
    a. Napomena: Proizvod se može koristiti u kombinaciji s dodatnim kompletom VIASURE za BD MAX test, a u tom slučaju odaberite opciju „Dual Master Mix Concentrated Lyophilized MM with Rehydration Buffer (Type 5)" (Dvostruka glavna smjesa koncentriran liofiliziran MM s rehidracijskim puferom - Tip 5).
6) U „Sample extraction parameters" (Parametri ekstrakcije uzorka) odaberite „User defined" (Korisnički definirani) i prilagodite volumen uzorka volumenu korištenog kliničkog uzorka plus 500 µl.
    a. Primjer: Ako ste pipetom prenijeli 200 µl respiratornog kliničkog uzorka u BD MAX TNA-3 Sample Buffer Tube, postavite parametar na 700 µl.
    b. Napomena: maksimalno možete postaviti 950 µl.
7) U „Ct Calculation" (Ct izračun) odaberite „Call Ct at Threshold Crossing" (Izračunaj Ct pri prelasku granice).
8) Na kartici „PCR settings" (Postavke za PCR) unesite sljedeće parametre: „Channel Settings" (Postavke kanala), „Gains" (Dobit) i „Threshold" (Granica) (Tablica 2).
    a. Napomena: Proizvod se može koristiti u kombinaciji s dodatnim kompletom VIASURE za BD MAX™, a u tom slučaju „PCR Settings" (Postavke za PCR) i „Test Steps" (Koraci testa) treba popuniti za položaje 2 (zeleni) i 4 (plavi).

Channel (Kanal)	Alias (Drugi naziv)	Gain (Dobit)	Threshold (Granica)	Ct Min (Ct minimalno)	Ct Max (Ct maksimalno)
475/520 (FAM)	SARS-CoV-2 S gene	60	100	0	40
530/565 (HEX)	IC	80	100	0	40
585/630 (ROX)	-	0	0	0	0
630/665 (Cy5)	-	0	0	0	0
680/715 (Cy5.5)	-	0	0	0	0

Tablica 2. Postavke za PCR.

Napomena: Preporučuje se postavljanje gore navedenih minimalnih graničnih vrijednosti za svaki kanal kao početnu točku, ali završne postavke mora odrediti krajnji korisnik tijekom tumačenja rezultata kako bi se osiguralo da granice spadaju u eksponencijalnu fazu krivulja fluorescencije i iznad bilo kojeg pozadinskog signala. Granična vrijednost različitih instrumenata može se razlikovati zbog različitih intenziteta signala.

9) Na kartici „PCR settings“ (Postavke za PCR) unesite i sljedeće parametre: „Spectral Cross Talk“ (Tablica 3)

		False Receiving Channel (Kanal s lažnim rezultatima)				
	Channel (Kanal)	475/520	530/565	585/630	630/665	680/715
Excitation Channel (Kanal za ekscitaciju)	475/520	-	0.0	0.0	0.0	0.0
	530/565	0.0	-	0.0	0.0	0.0
	585/630	0.0	0.0	-	0.0	0.0
	630/665	0.0	0.0	0.0	-	0.0
	680/715	0.0	0.0	0.0	0.0	-

Tablica 3. Parametri spektralnog preklapanja signala

10) Na kartici „Test Steps“ (Koraci testa) unesite protokol za PCR (Tablica 4).

Step Name (Naziv koraka)	Profile Type (Vrsta profila)	Cycles (Ciklusi)	Time (s) (Vrijeme (s))	Temperature (Temperatura)	Detect (Detekcija)
Reverse transcription (Reverzna transkripcija)	Hold (Čekanje)	1	900	45°C	-
Initial denaturation (Početna denaturacija)	Hold (Čekanje)	1	120	98°C	-
Denaturation and Annealing/Extension (Data collection) (Denaturacija i sparivanje/produljenje (prikupljanje podataka))	2-Temperature (2-temperatura)	45	10	95°C	-
			58	60°C	✓

Tablica 4. Protokol za PCR.

11) Kliknite na gumb „Save Test“ (Spremi test).

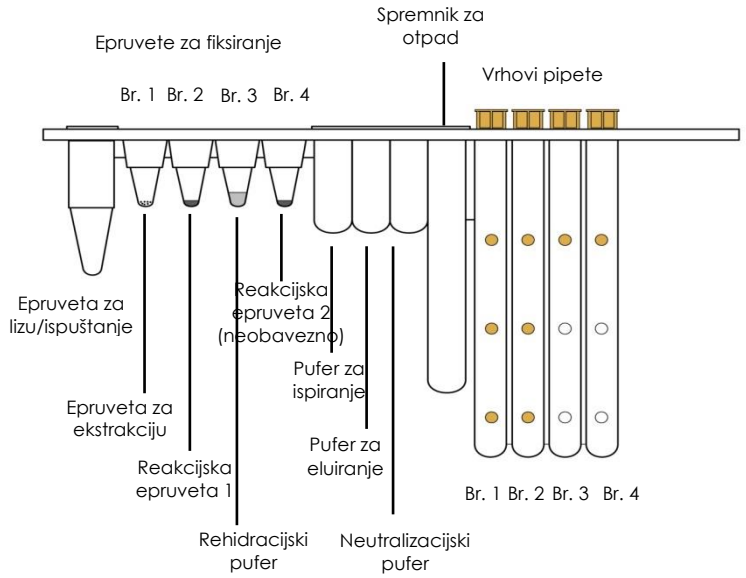
8.3.2. Postavljanje BD MAX™ stalka

1) Za svaki uzorak koji treba testirati izvadite pojedinačne trake s reagensima u bloku iz kompleta BD MAX™ ExK TNA-3 kit. Lagano udarite svaku traku od čvrstu površinu kako biste osigurali da se sve tekućine nalaze na dnu epruveta i stavite ih na stalke za uzorke BD MAX™ sustava.
2) Izvadite potrebni broj BD MAX™ ExK TNA epruveta za ekstrakciju (B4) (bijela folija) iz njihove zaštitne vrećice. Postavite epruvetu/e za ekstrakciju (bijela folija) u njihove odgovarajuće položaje u TNA traku (položaj 1 s oznakom u boji na stalku. Pogledajte Sliku 1). Istisnite višak zraka i zatvorite vrećicu patentnim zatvaračem.
3) Odredite i razdvojite odgovarajući broj reakcijskih epruveta VIASURE *SARS-CoV-2 S gene* reaction tube (zelena folija) i postavite ih u njihove odgovarajuće položaje na traci (položaj 2, označen zelenom bojom na stalku. Pogledajte Sliku 1).
    a. Istisnite višak zraka i zatvorite aluminijske vrećice patentnim zatvaračem.
    b. U cilju obavljanja pravilnog postupka rehidracije pazite da se liofilizirani proizvod nalazi na dnu epruvete te da ne prianja za gornji dio epruvete ili za zatvarač od folije.

i. Napomena: Ako odaberete oblik „Dual Master Mix Concentrated Lyophilized MM with rehydration Buffer (Type 5)“ (Dvostruka glavna smjesa koncentriran liofiliziran MM s rehidracijskim puferom - Tip 5) (odjeljak 8.3.1), odredite i razdvojite odgovarajući broj dodatnih reakcijskih epruveta VIASURE reaction tube (drugačija folija) i postavite ih u njihove odgovarajuće položaje na traci (položaj 4, označen plavom bojom na stalku. Pogledajte Sliku 1). Istisnite višak zraka i zatvorite aluminijske vrećice patentnim zatvaračem.

4) Izvadite odgovarajući broj epruveta Rehydration Buffer tubes (narančasta folija) i postavite ih u njihove odgovarajuće položaje na traci (položaj 3, nije obojen na stalku. Pogledajte Sliku 1). Istisnite višak zraka i zatvorite vrećicu patentnim zatvaračem.
   a. U cilju osiguravanja pravilnog prijenosa pazite da se tekućina nalazi na dnu epruvete te da ne prianja za gornji dio epruvete ili za zatvarač od folije.

Slika 1. BD MAX™ TNA traka s reagensima (TNA) iz kompleta BD MAX™ ExK TNA-3 kit.



8.3.3. Postavljanje BD MAX™ instrumenta

1) Odaberite karticu „Work List“ (Radni popis) na zaslonu „Run“ (Pokreni) na BD MAX™ sustavu s verzijom softvera v4.50A ili novijom.
2) S padajućeg izbornika „Test“ odaberite VIASURE *SARS-CoV-2 S gene* (ako test nije već kreiran, pogledajte odjeljak 8.3.1).
3) Odaberite odgovarajući broj serije kompleta (naveden na vanjskoj kutiji kompleta za ekstrakciju kojeg koristite) s padajućeg izbornika (neobavezno).
4) Unesite identifikacijski broj epruvete za uzorak s puferom u prozor „Sample tube“ (Epruveta s uzorkom) na kartici „Worklist“ (Radni popis) skeniranjem crtičnog koda pomoću sekenera ili ručnim unošenjem broja.
5) Popunite polje „Specimen/Patient ID“ (ID uzorka/bolesnika) i/ili prozor „Accession“ (Pristup) na kartici „Worklist“ (Radni popis) i kliknite na gumb „Save“ (Spremi). Nastavite s postupkom dok ne unesete sve

epruvete za uzorak s puferom. Pazite da se ID uzorka/bolesnika i epruvete za uzorak s puferom podudaraju.

6) Stavite pripremljenu epruvetu za uzorak s puferom na BD MAX™ stalak/e.
7) Stavite stalak/e u BD MAX™ sustav (stalak A se nalazi lijevo u odnosu na BD MAX™ sustav, dok se stalak B nalazi na desnoj strani).
8) Stavite potrebni broj BD MAX™ PCR patrona u BD MAX™ sustav.
9) Zatvorite vrata BD MAX™ sustava.
10) Kliknite „Start Run" (Pokreni postupak) da započnete postupak.

8.3.4. BD MAX™ izvješće

1) U glavnom izborniku kliknite na gumb „Results" (Rezultati).
2) Kliknite dvaput na svoj postupak na popisu ili pritisnite gumb „view" (prikaz).
3) Kliknite na „Print" (Ispiši) i odaberite: "Run Details, Test Details and Plot…" (Podaci o postupku, podaci o testu i grafikon...)
4) Kliknite na gumb „Print or Export" (ispiši ili izvezi) na zaslonu „Run Reports" (Izvješća o postupku)

9. Tumačenje rezultata

Detaljne upute za analiziranje podataka potražite u Korisničkom priručniku za BD MAX™ sustav.

Analizu podataka obavlja BD MAX™ softver prema uputama proizvođača. BD MAX™ softver kreira izvješće o Ct vrijednostima i amplifikacijskim krivuljama za svaki kanal detekcije svakog uzorka testiranog na sljedeći način:

- Ct vrijednost 0 znači da softver nije izračunao Ct vrijednost s navedenom granicom (pogledajte Tablicu 2). Amplifikacijska krivulja uzorka koja pokazuje „0" za Ct vrijednost mora se provjeriti ručno.

- Ct vrijednost -1 znači da nije došlo do amplifikacijskog procesa.

- Svaku drugu Ct vrijednost treba tumačiti u vezi s amplifikacijskom krivuljom i prema smjernicama za tumačenje uzoraka navedenim u Tablici 5.

Provjerite unutarnji kontrolni signal da potvrdite funkcionira li amplifikacijska smjesa ispravno. Pored toga, provjerite da nema izvješća o kvaru BD MAX™ sustava.

-Pomoću sljedeće tablice očitajte i analizirajte rezultate:

SARS-CoV-2 *S* gene (475/520)	Unutarnja kontrola (530/565)	Tumačenje
-	+	**RNK SARS-CoV-2 *S* gena nije otkrivena**
+	+/-	**RNK SARS-CoV-2 *S* gena je otkrivena**
-	-	**Neriješen (UNR) rezultat dobiven u prisutnosti inhibitora reakcije PCR-a ili kada dođe do nekog općeg problema (koji nije pokriven šifrom o pogrešci) pri obradi uzorka i/ili amplifikaciji.**
IND	IND	**Rezultat testa nije moguće utvrditi (IND). Zbog kvara BD MAX™ sustava. Rezultat testa prikazan u slučaju kvara instrumenta povezanog sa šifrom o pogrešci.**
INC	INC	**Nepotpun rezultat testa (INC). Zbog kvara BD MAX™ sustava. Rezultat testa prikazan u slučaju neuspješnog obavljanja postupka.**

Tablica 5. Tumačenje rezultata

+: Došlo je do amplifikacije

-: Nije došlo do amplifikacije

Uzorak se smatra pozitivnim ako je dobivena Ct vrijednost manja od 40. Unutarnja kontrola može ili ne mora pokazati amplifikacijski signal, jer veliki broj kopija ciljne nukleinske kiseline može izazvati preferencijalnu amplifikaciju specifičnih nukleinskih kiselina umjesto unutarnje kontrole. U tim slučajevima nije potrebna detekcija unutarnje kontrole.

Uzorak se smatra negativnim ako ne pokazuje amplifikacijski signal u sustavu detekcije, ali je unutarnja kontrola pozitivna. Inhibiranje reakcije PCR-a može se isključiti amplifikacijom unutarnje kontrole.

U slučaju neriješenih rezultata (UNR), ako signal unutarnje kontrole nije prisutan u negativnom uzorku, preporučuje se ponavljanje testa razrjeđivanjem uzorka u omjeru 1:10 kako bi se provjerilo postojanje eventualnih problema s inhibicijom.

Rezultate testa treba procijeniti zdravstveni djelatnik uzimajući u obzir anamnezu, kliničke simptome i druge dijagnostičke testove.

10. Ograničenja testa

- Rezultate testa treba procijeniti zdravstveni djelatnik uzimajući u obzir anamnezu, kliničke simptome i druge dijagnostičke testove.
- Iako se ovaj test može koristiti s drugim vrstama uzoraka, validiran je s nazofaringealnim/orofaringealnim brisovima prikupljenima u VTM-u.
- Kvaliteta testa ovisi o kvaliteti uzorka; mora se ekstrahirati odgovarajuća nukleinska kiselina iz respiratornih uzoraka.
- Izuzetno niske razine ciljnog elementa ispod razine detekcije mogu biti detektirane, no moguće je da se rezultati neće moći ponoviti.

- Postoji mogućnost pojave lažnih pozitivnih rezultata zbog unakrsne kontaminacije izazvane uzorcima suspektnima na SARS-CoV-2 koji sadrže visoke koncentracije ciljne RNK ili kontaminacije izazvane proizvodima koji sadrže PCR iz ranijih reakcija.
- Specifična kombinacija početnice i sonde za detekciju gena *S* gene korištena u kompletu VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit dizajniranom za detekciju SARS-CoV-2, ne pokazuju značajne kombinirane homologije s ljudskim genomom, ljudskom mikroflorom, SARS-CoV ili drugim koronavirusima, što bi moglo rezultirati predvidljivim lažno pozitivnim rezultatima.
- Lažni negativni rezultati mogu se pojaviti uslijed nekoliko čimbenika i njihovih kombinacija, uključujući:
    - Nepravilno prikupljanje, transport, pohrana i/ili metode rukovanja.
    - Nepravilni postupci obrade (uključujući ekstrakciju RNK).
    - Propadanje virusni RNK tijekom otpremanja/pohrane i/ili obrade uzorka.
    - Mutacije ili polimorfizam u regijama za vezivanje početnice ili sonde mogu utjecati na detekciju novih ili nepoznatih varijanti SARS-CoV-2.
    - Virusno opterećenje u uzorku ispod granice detekcije testa.
    - Prisutnost inhibitora RT-qPCR ili drugih vrsta ometajućih tvari. Nisu procijenjeni utjecaji cjepiva, antivirusnih terapija, antibiotika, kemoterapijskih sredstva ili imunosupresiva korištenih za prevenciju COVID-19 ili tijekom liječenja infekcije.
    - Nepridržavanje uputa za uporabu i postupka testiranja.
- Negativni rezultati ne isključuju infekciju SARS-CoV-2 i ne smiju se koristiti kao jedini temelj za odlučivanje o liječenju ili pružanju druge zdravstvene skrbi bolesniku. Nisu određene optimalne vrste uzoraka i izračun vremena vršnih razina virusa tijekom infekcija čiji je uzrok SARS-CoV-2. Za detekciju virusa može biti potrebno prikupljanje više uzoraka (vrste i vremenske točke) od istog bolesnika.
- Ako su dijagnostički testovi za ostale respiratorne bolesti negativni, a klinička slika bolesnika te epidemiološke informacije sugeriraju da je moguća infekcija SARS-CoV-2, potrebno je razmotriti mogućnost lažnog negativnog rezultata i ponovnog testiranja bolesnika.
- Negativni rezultat ne isključuje prisutnost RNK virusa SARS-CoV-2 u kliničkom uzorku. Ako kliničko promatranje, povijest bolesnika i epidemiološke informacije sugeriraju infekciju COVID-19, potrebno je razmotriti ponovno testiranje na uzorku većeg volumena.
- U slučaju neriješenih, neutvrdivih ili nepotpunih rezultata primjenom VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit potrebno je ponovno testiranje. Neriješeni rezultati mogu biti posljedica prisutnosti inhibitora u uzorku ili nepravilne rehidracije epruvete s liofiliziranom reakcijskom smjesom. Ako postoji kvar na instrumentu, dobivaju se neutvrdivi ili nepotpuni rezultati.

11. Kontrola kvalitete

Komplet VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit sadrži unutarnju kontrolu (IC) u svakoj reakcijskoj epruveti pomoću koje se potvrđuje pravilno provođenje tehnike.

12. Radne karakteristike

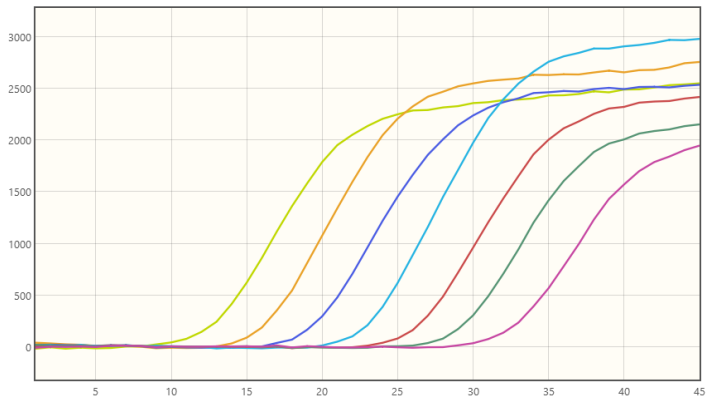
12.1. Klinička osjetljivost i specifičnost

Klinički učinak kompleta VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit testiran je na 4 nukleinske kiseline izolirane iz pozitivnih nazofaringealnih i/ili orofaringealnih brisova prikupljenima u VTM-u i na 15 respiratornih uzoraka (nazofaringealnih i/ili orofaringealnih brisova u VTM-u) od bolesnika s kliničkom sumnjom na bolest COVID-19 ili druge slične respiratorne bolesti. Pronađeno je 4 pozitivnih uzoraka na SARS-CoV-2 i ti su rezultati bili u skladu s testom PCR koji je razvijen na temelju početnica i sondi CDC-a iz Kine za detekciju SARS-CoV-2.
Osim toga, 4 pozitivnih uzoraka na SARS-CoV-2 potvrđeno je primjenom metode molekularne detekcije španjolskog nacionalnog referentnog centra (Instituto de Salud Carlos III - ISCIII))(protokol "2019-nCoV by real-time RT-PCR" koji je predložio Charité (Berlin), s izmjenama).
Zaključno, rezultati pokazuju visoku osjetljivost i specifičnost detekcije SARS-CoV-2 pomoću kompleta VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit.

12.2. Analitička osjetljivost

Granica detekcije kompleta VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit iznosi ≥24 kopija cDNK po reakciji (cp/rxn) s pozitivnom stopom od ≥95%.

Slika 2. Serija razrjeđivanja predloška SARS-CoV-2 *S* gena ($2{,}4*10^7$-$2{,}4*10^1$ cp/rxn) analizirana na BD MAX™ System (475/520 (FAM) kanal).



12.3. Analitička specifičnost

Specifičnost testa SARS-CoV-2 *S* gene potvrđena je testiranjem panela koji se sastoji od različitih mikroorganizama koji predstavljaju najčešće respiratorne patogene. Nije zabilježena unakrsna reaktivnost između bilo kojih od sljedećih testiranih mikroorganizama:

Testiranje unakrsne reaktivnosti					
Ljudski adenovirus tipovi 1-5, 8, 15, 31, 40 i 41	-	Influenza A/California/7/2009(H1N1)pdm09-like virus	-	*Legionella dumoffii*	-
Ljudski bokavirus	-	Influenza A/Michigan/45/2015 (H1N1)pdm09 virus	-	*Legionella longbeachae*	-
Bordetella bronchiseptica	-	Influenza A/Singapore/GP1908/2015, IVR-180 (H1N1)pdm09 virus	-	*Legionella micdadei*	-
Bordetella holmesii	-	Influenza A/Victoria/210/2009 (H3N2) virus	-	*Legionella pneumophila*	-
Bordetella parapertussis	-	Influenza A/Thüringen/5/17 (H3N2) virus	-	Ljudski metapneumovirus A i B	-
Bordetella pertussis	-	Influenza A/Switzerland/9715293/2013 (H3N2) virus	-	*Moraxella catarrhalis*	-
Chlamydia caviae	-	Influenza A/Hong Kong/4801/2014, NYMC X-263B (H3N2) virus	-	*Mycoplasma pneumoniae*	-
Chlamydia psittaci genotip A i C	-	Influenza A/ South Australia/55/2014, IVR-175 (H3N2) virus	-	*Mycobacterium tuberculosis*	-
Chlamydophila pneumoniae	-	Influenza A/DE-SH/Reiherente/AR8444/ 2016 (H5N8) virus	-	Virusi ljudske parainfluence tipa 1, 2, 3 i 4	-
Ljudski koronavirus 229E, OC43, NL63 i HKU1	-	Influenza A/Anhui/1/2013 (H7N9) virus	-	*Pneumocytis jirovecii* tip A1 i g885652	-
MERS koronavirus	-	Influenza B/Brisbane/60/2008-like virus	-	Ljudski rinovirus tip C	-
SARS koronavirus soj Frankfurt 1	-	Influenza B/Florida/04/06 virus	-	*Staphylococcus aureus* podsoj *aureus*	-
Haemophilus influenzae MinnA	-	Influenza B/Phuket/3073/2013 virus	-	*Streptococcus pneumoniae*	-
Influenza A/New Caledonia/20/99(H1N1) virus	-	*Legionella bozemanii*	-	Respiratorni sincicijski virus (RSV) A i B	-

Tablica 6. Referentni patogeni mikroorganizmi korišteni u ovom ispitivanju.

12.4. Analitička reaktivnost

Reaktivnost kompleta VIASURE *SARS-CoV-2 S gene* Real Time PCR Detection Kit procijenjena je u odnosu na RNK iz soja ljudskog 2019-nCoV BetaCoV/Germany/BavPat1/2020 p.1, iz soja ljudskog 2019-nCoV 2019-nCoV/Italy-INMI1, kontrole sintetičkog RNK za dvije varijante virusa SARS-CoV-2: MT007544.1 (SARS-CoV2 izolat Australia/VIC01/2020) i MN908947.3 (SARS-CoV-2 izolat Wuhan-Hu-1), i pokazala je pozitivne rezultate.

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14. Symbols for IVD components and reagents/Simboli za IVD komponente i reagense

IVD	*In vitro* diagnostic device *In vitro* dijagnostički uređaj		Keep dry Čuvati na suhom		Use by Rok valjanosti		Manufacturer Proizvođač	LOT	Batch code (Lot) Šifra serije
	Consult Instructions for Use Pogledajte upute za upotrebu		Temperature limitation Ograničenje temperature	Σ	Contains sufficient for <n> test Sadržaj dovoljan za <n> test(ova)	DIL	Sample diluent Razrjeđivač uzorka	REF	Catalognumber Kataloški broj

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IVD

Real Time PCR Detection Kits

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